

**INRAe**



## Metabarcoding : the main steps - Part 1

Clarissee Lemonnier



## Summary

The main steps of metabarcoding

Selection of the barcode

Reference databases

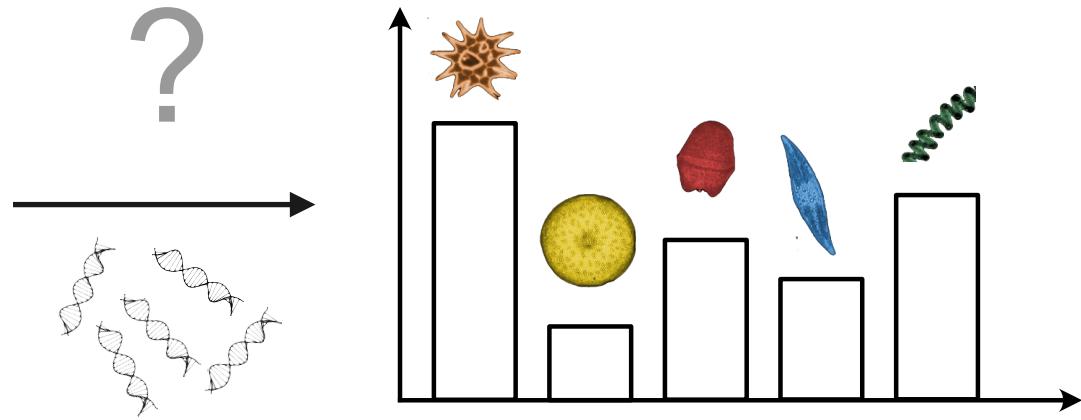


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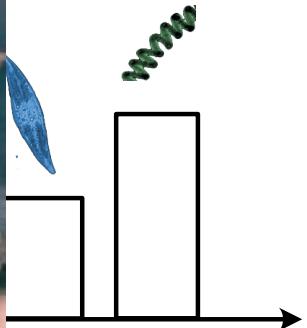
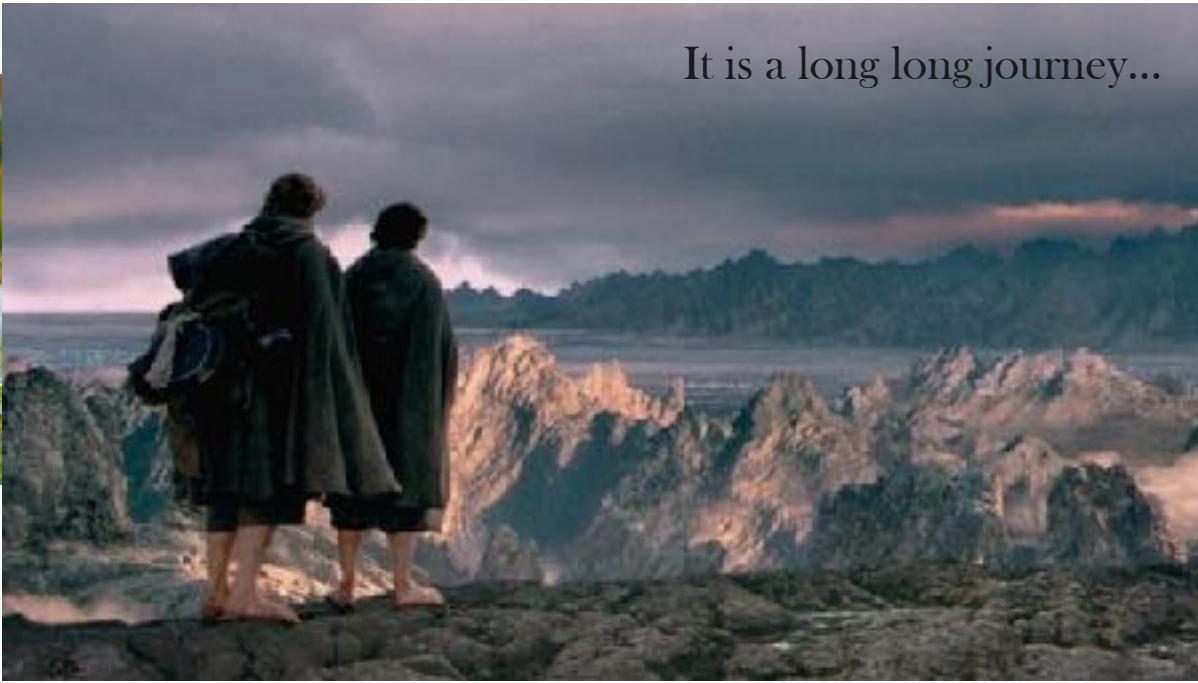
## Metabarcoding steps



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## Metabarcoding steps



## Metabarcoding steps

*Sampling*



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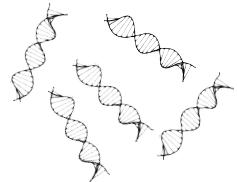
[www.biolaweb.com](http://www.biolaweb.com)

## Metabarcoding steps

*Sampling*



*DNA extraction*



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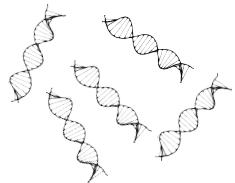
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## Metabarcoding steps

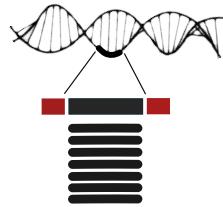
*Sampling*



*DNA extraction*



*Barcode amplification*



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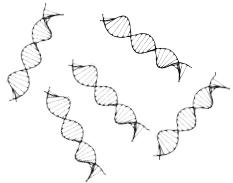
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## Metabarcoding steps

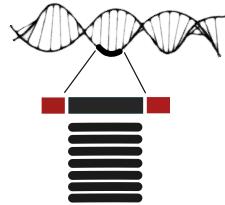
*Sampling*



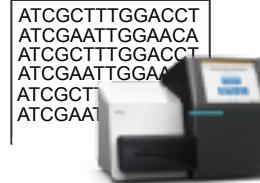
*DNA extraction*



*Barcode amplification*



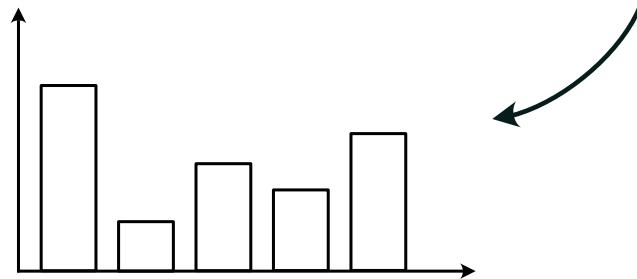
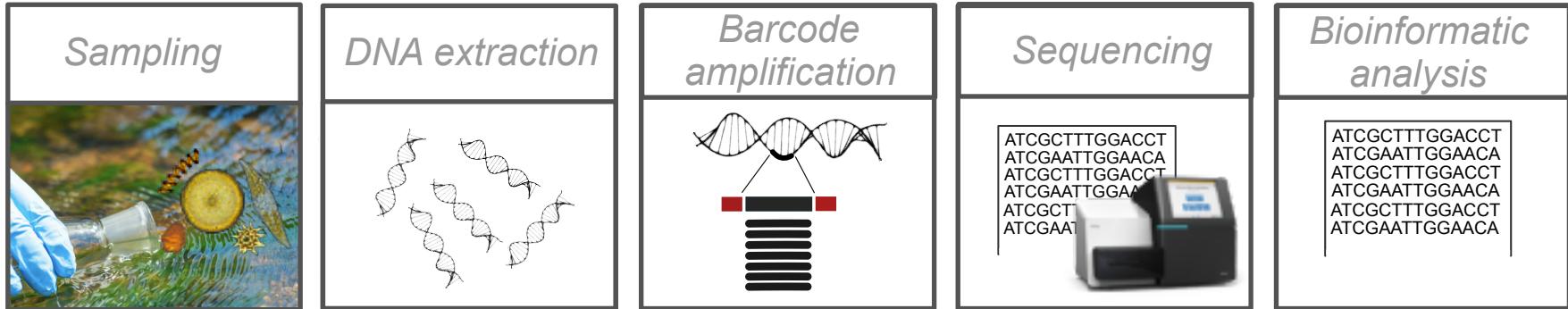
*Sequencing*



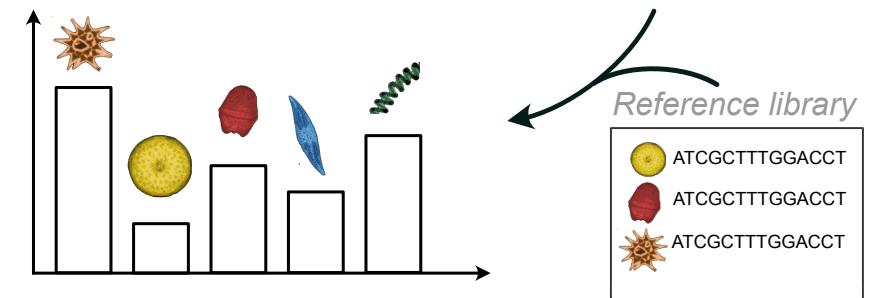
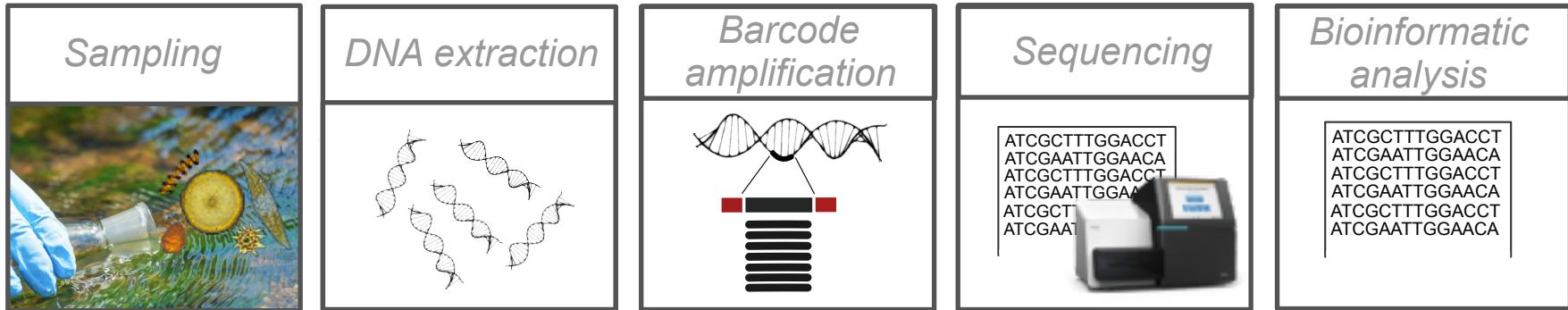
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## Metabarcoding steps



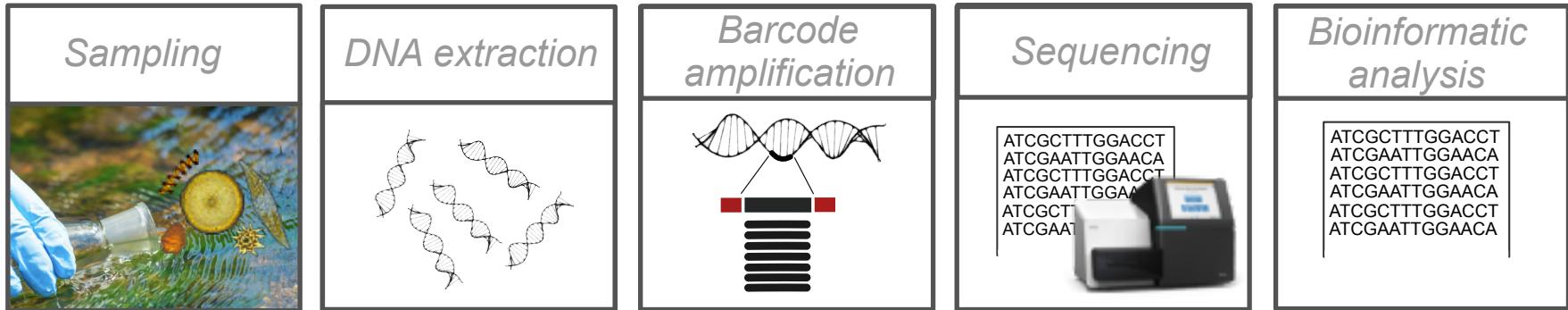
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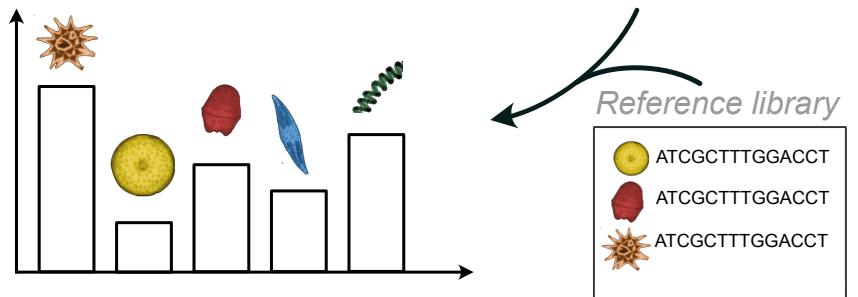
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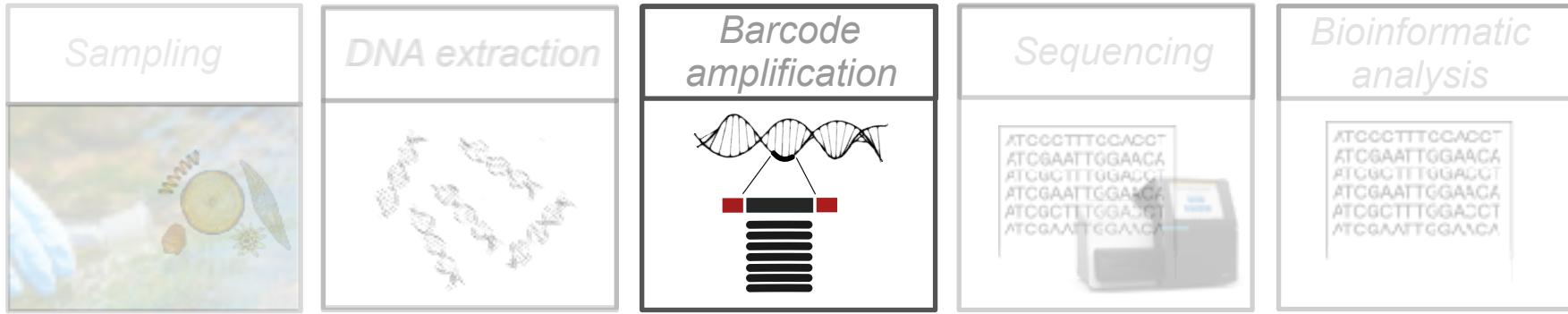
## Metabarcoding steps



**An informed choice at each step is required to get a reliable result at the end**



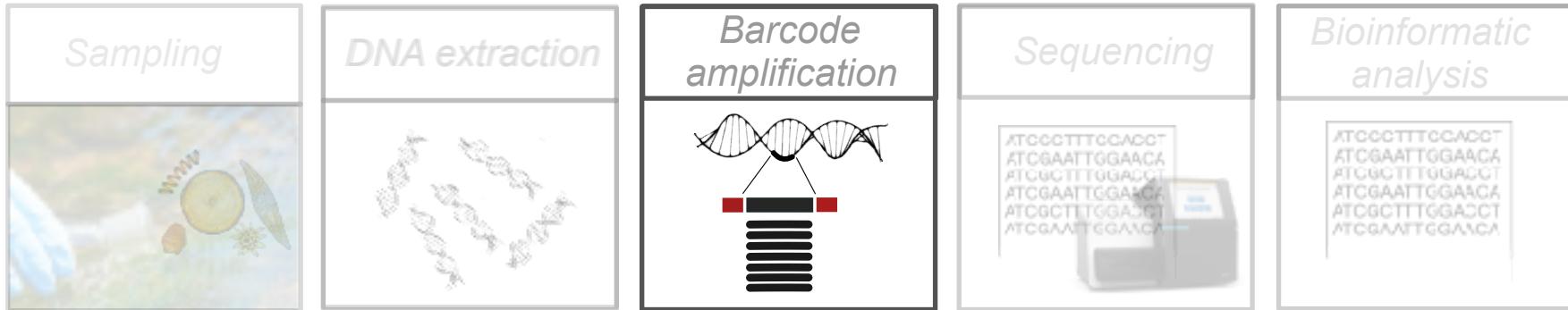
## Metabarcoding steps



### Barcode selection



## Metabarcoding steps



Recovery of all species



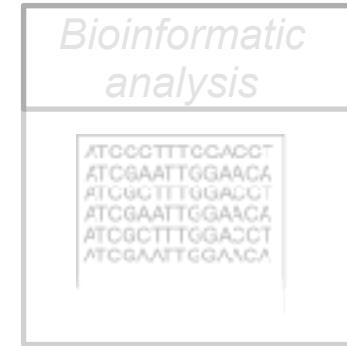
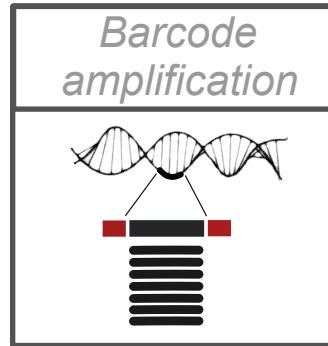
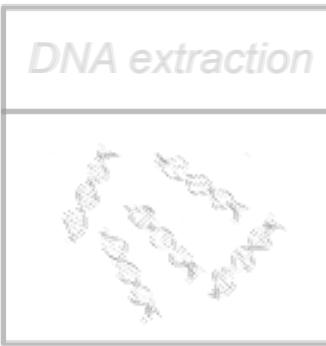
*Barcode selection*



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## Metabarcoding steps

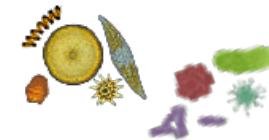


**Recovery of all species**

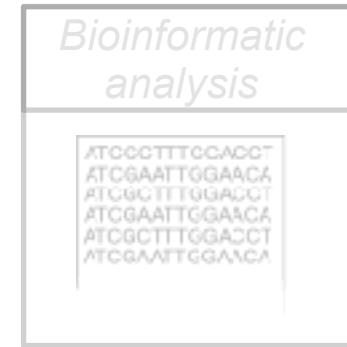
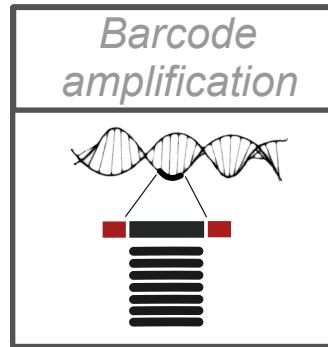


**Barcode selection**

**Specificity**



## Metabarcoding steps



Recovery of all species



*Barcode selection*

Taxonomic resolution and identification

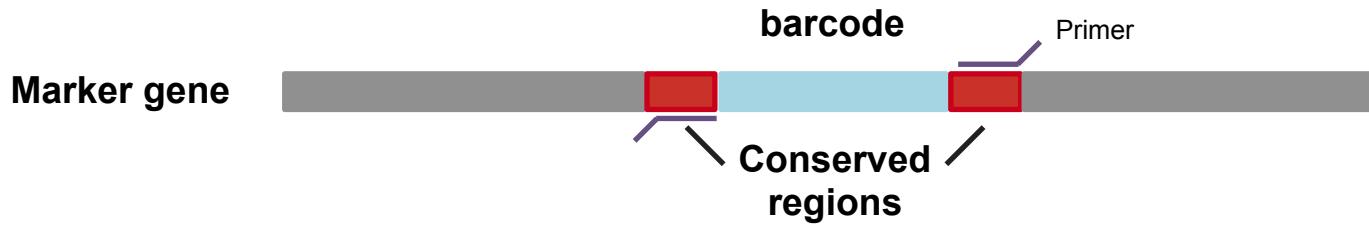
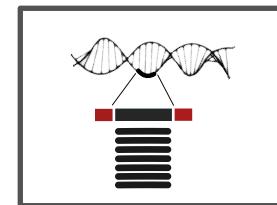
Specificity



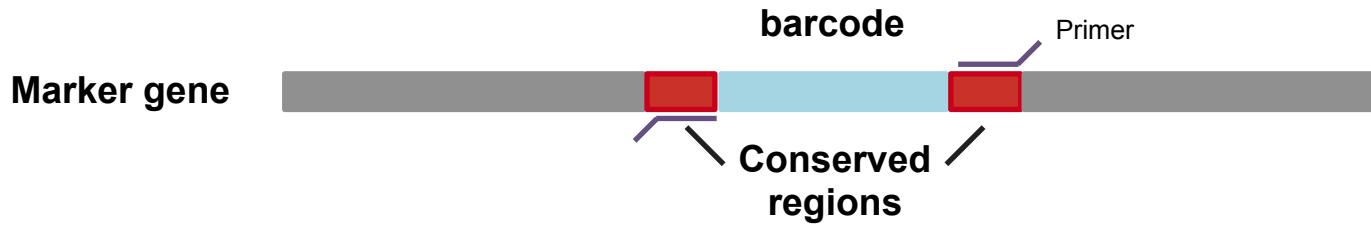
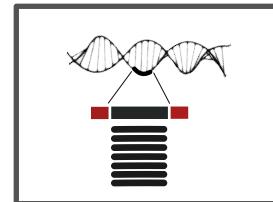
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## Barcode selection - a fundamental criteria



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1

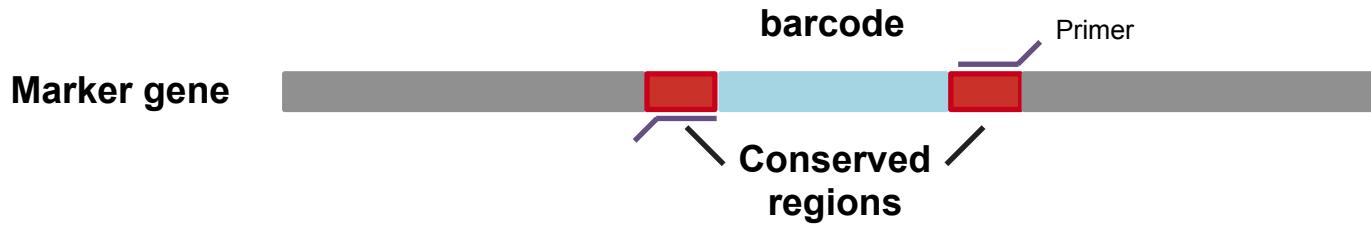
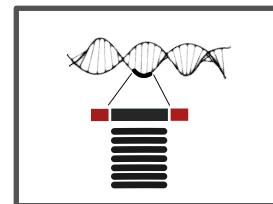
Universal



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## Barcode selection - a fundamental criteria



1 Universal

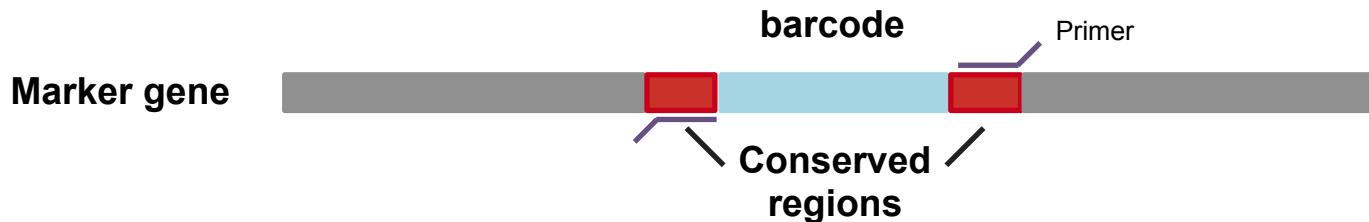
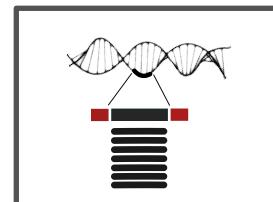
2 Conserved regions



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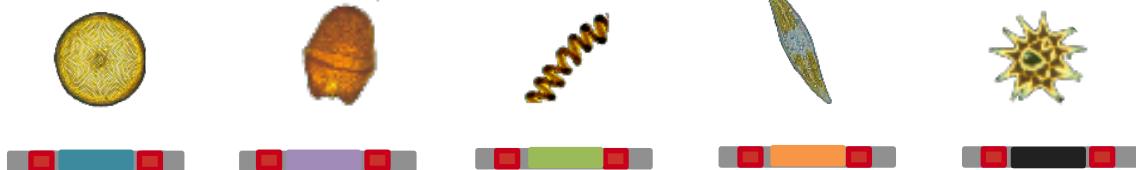
## Barcode selection - a fundamental criteria



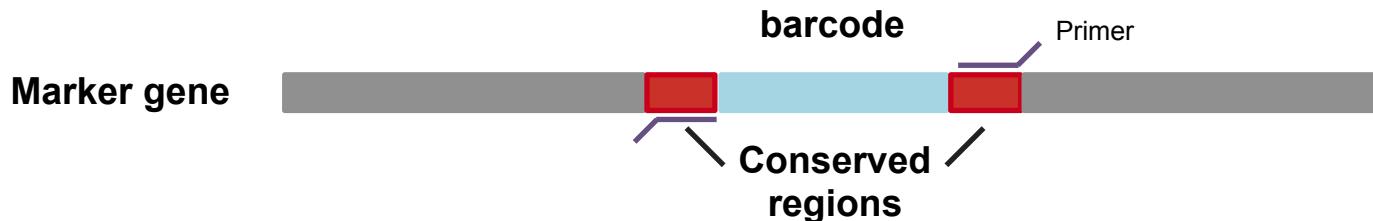
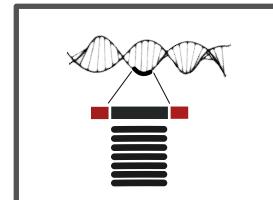
1 Universal

2 Conserved regions

3 Variable enough



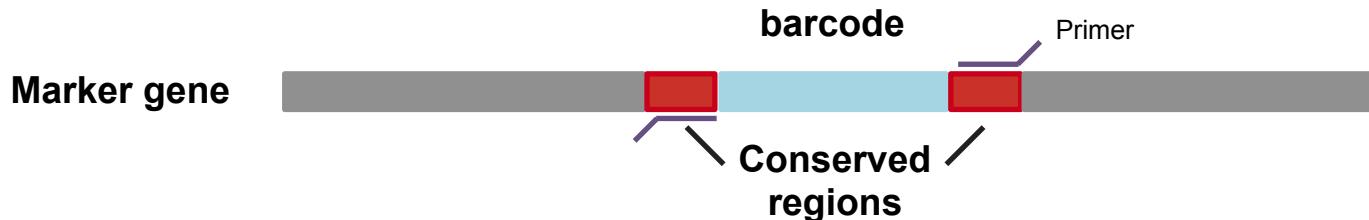
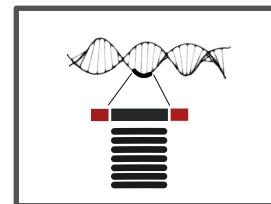
## Barcode selection - a fundamental criteria

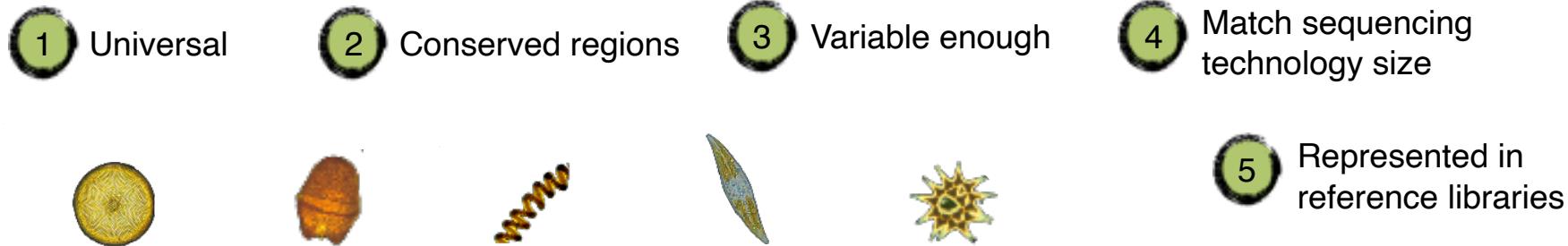


- 1 Universal
- 2 Conserved regions
- 3 Variable enough
- 4 Match sequencing technology size



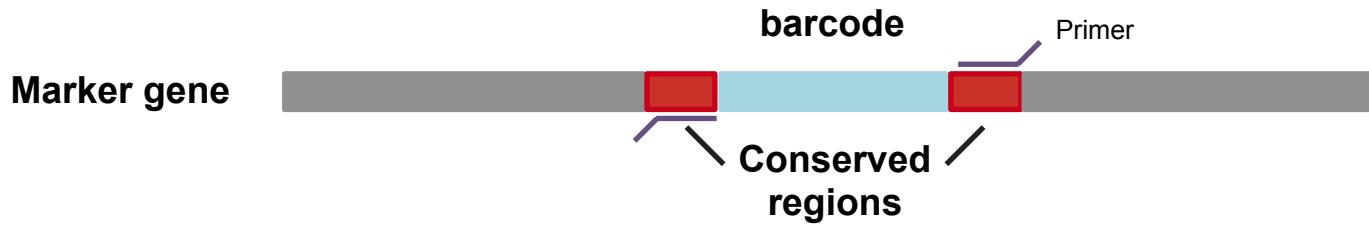
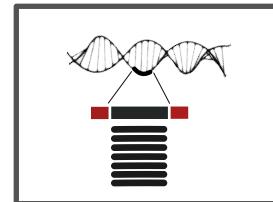
## Barcode selection - a fundamental criteria



- 1 Universal
  - 2 Conserved regions
  - 3 Variable enough
  - 4 Match sequencing technology size
  - 5 Represented in reference libraries
- 



## Barcode selection - a fundamental criteria



1 Universal

2 Conserved regions

3 Variable enough

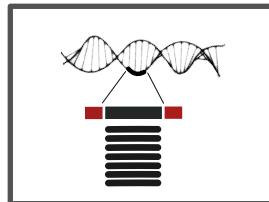
4 Match sequencing technology size

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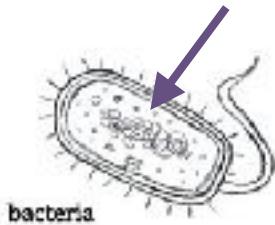
What are the classical marker genes used in metabarcoding?



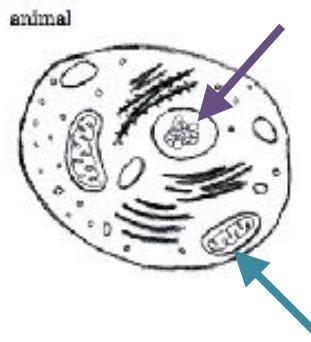
## Classical marker genes - an overview



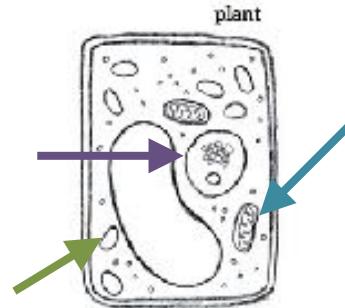
- A marker gene can be present in all organisms genomes



Genomic DNA



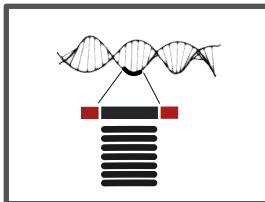
Mitochondrial DNA



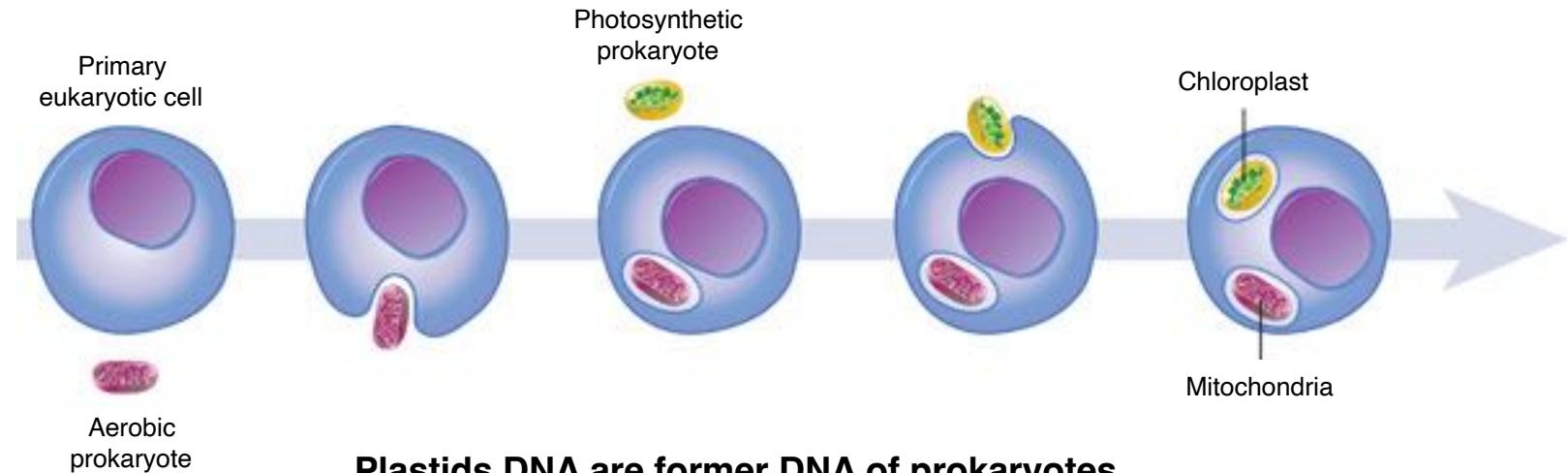
Chloroplast DNA



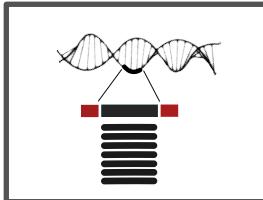
## Classical marker genes - an overview



- A marker gene can be present in all organisms genomes



# Classical marker genes - an overview

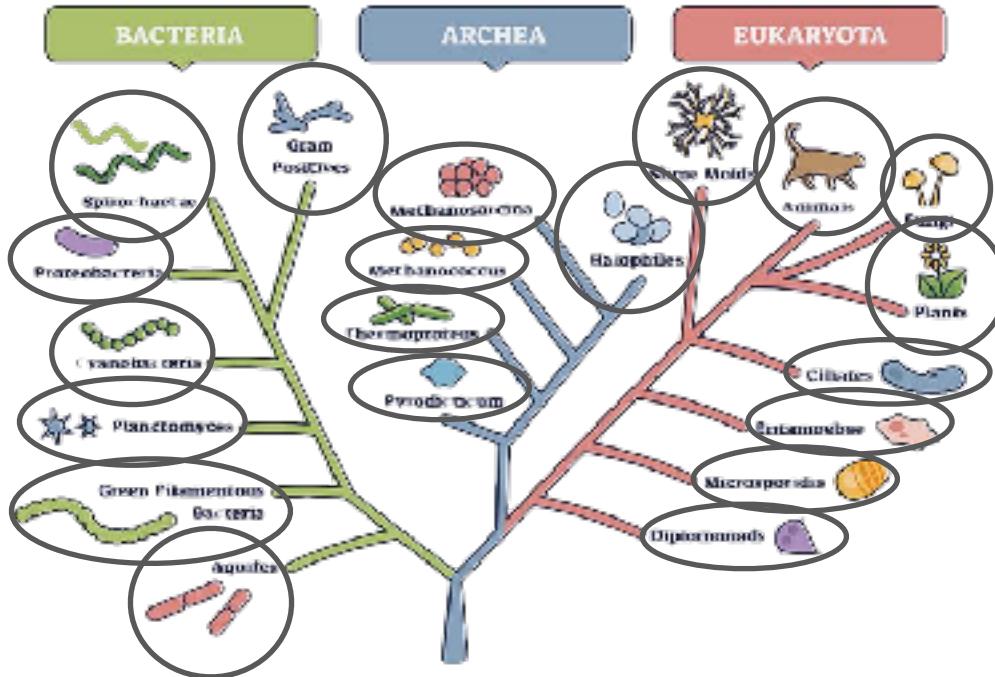


Ribosomal proteins

Nuclear

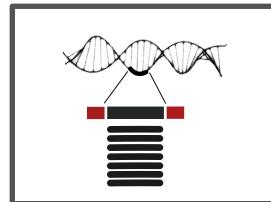
Mitochondrial

Chloroplast



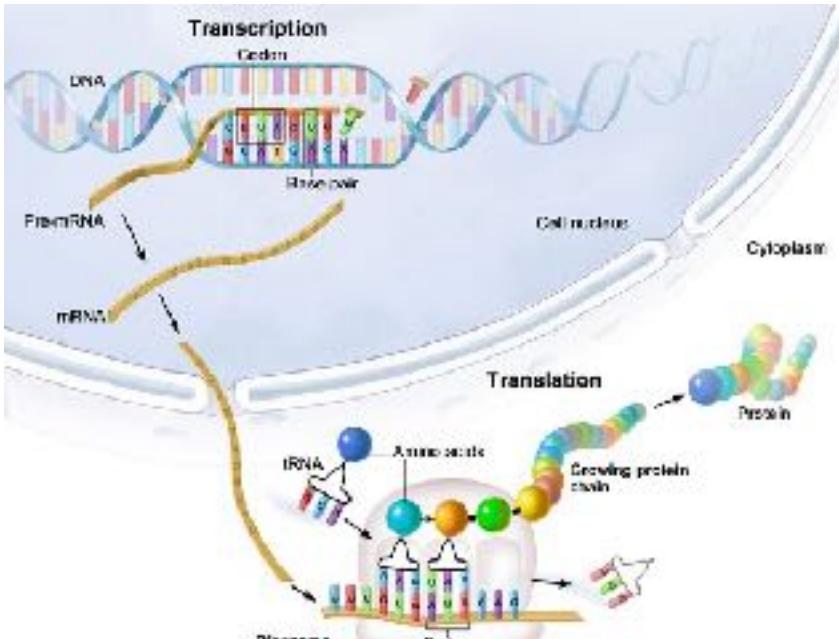
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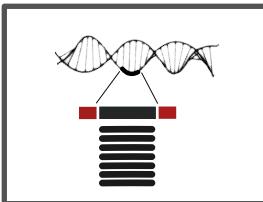
## Ribosomal genes

- Code for the large and small units of ribosomes which are responsible of the translation of mRNA into proteins
- Present in the nuclear DNA of all living organisms
- Present in the plastid DNA (mitochondria, chloroplasts)



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EU Project: BioLaweb

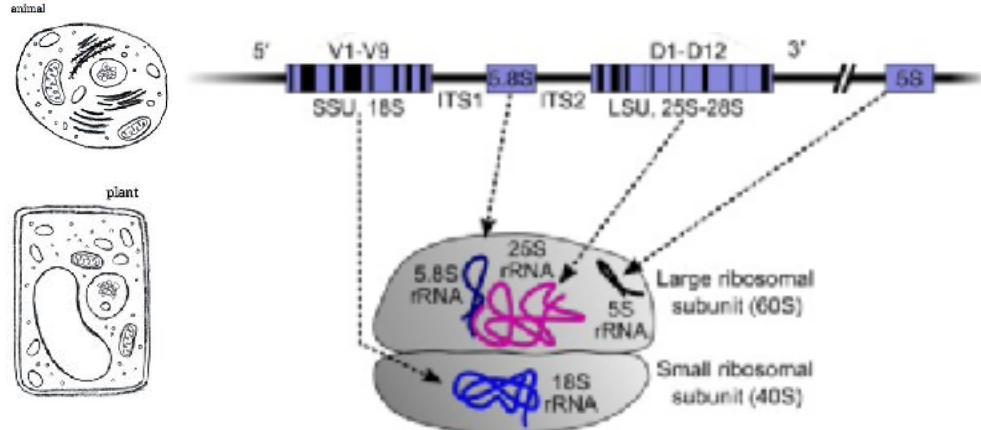


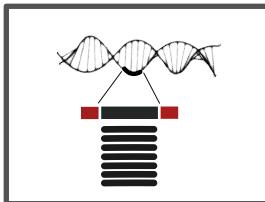


## Ribosomal genes



Genes organized in an operon (*i.e.* clustered in the genome)

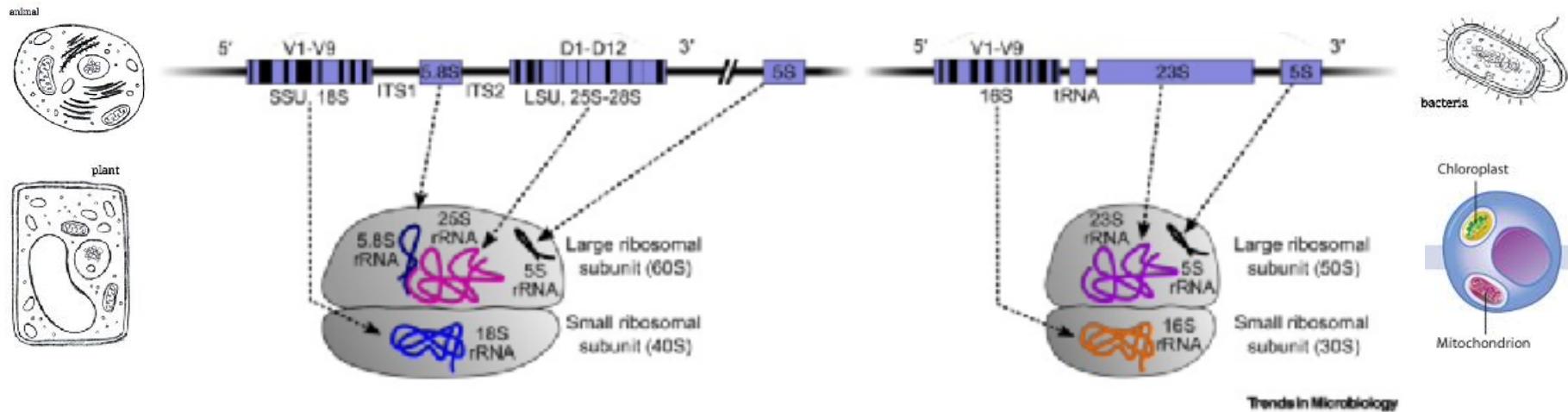


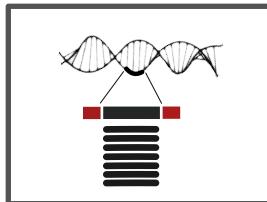


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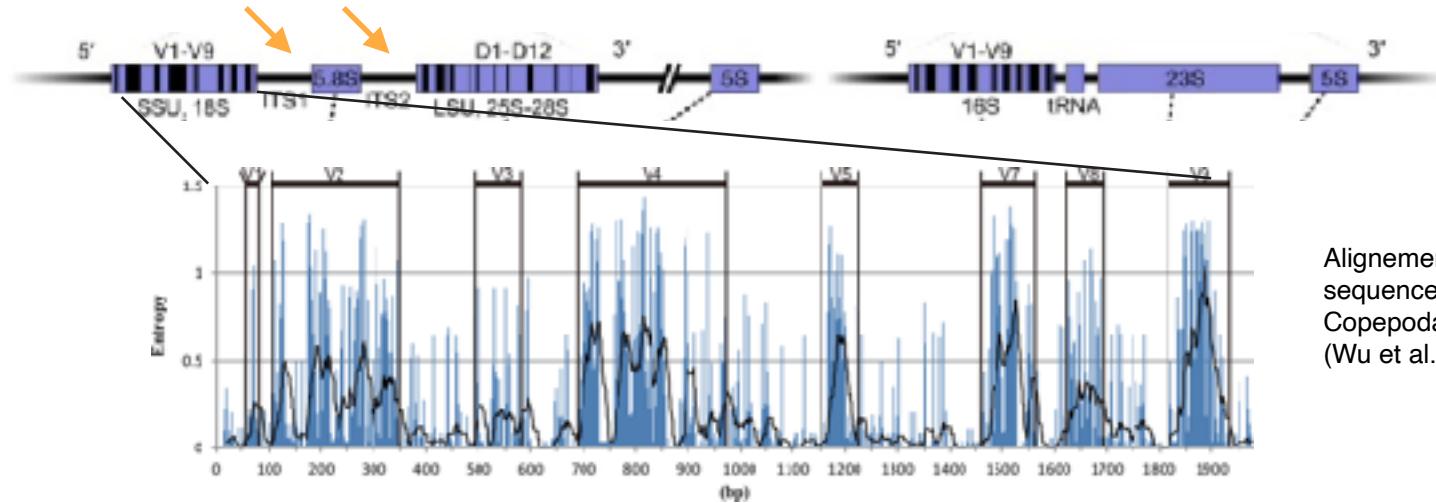




## Ribosomal genes

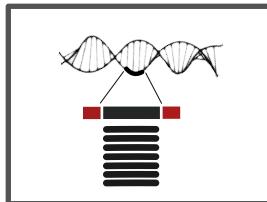


Alternance of hyper variable and conserved regions + Internal Transcribe spacer (ITS)



Alignement of 18S  
sequences from 192  
Copepoda species  
(Wu et al., 2015)

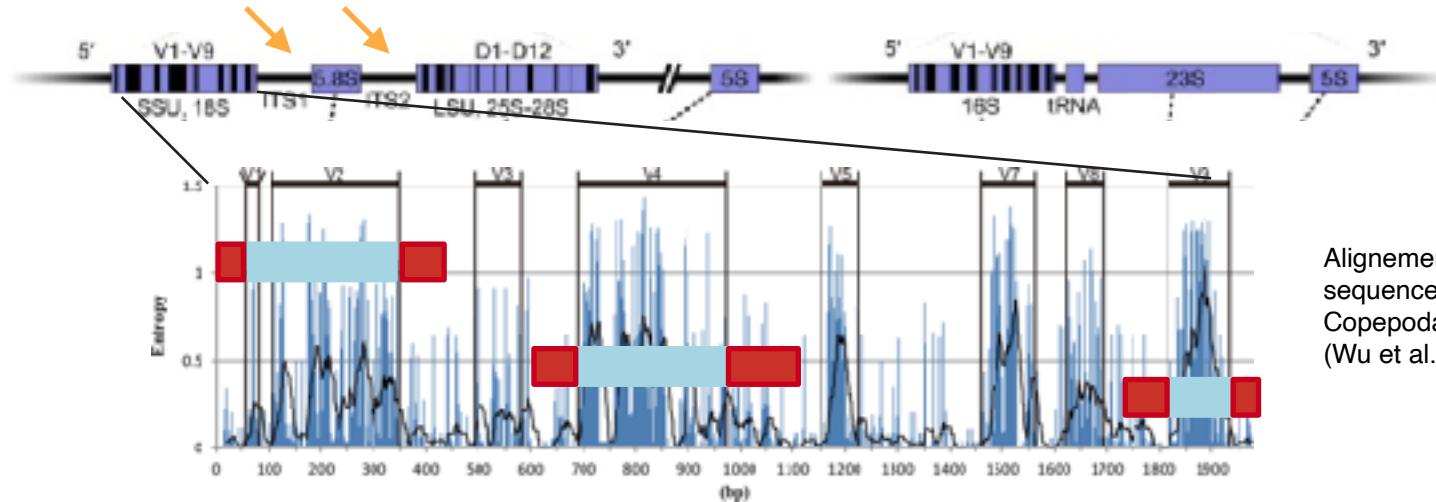




## Ribosomal genes



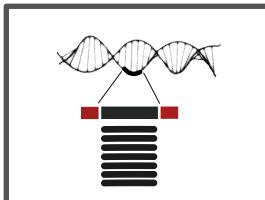
Alternance of hyper variable and conserved regions + Internal Transcribe spacer (ITS)



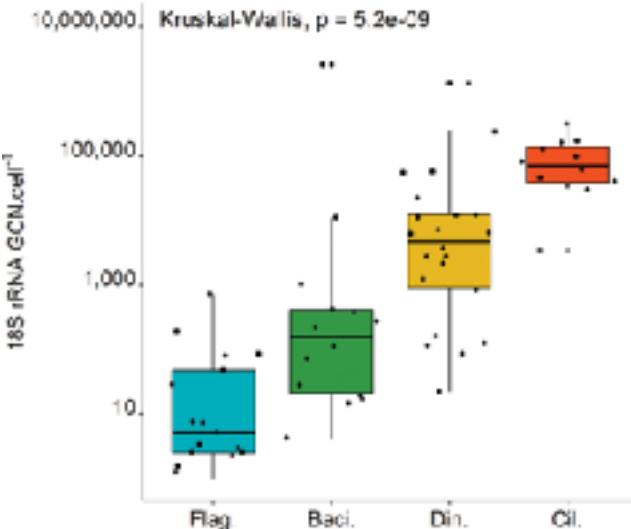
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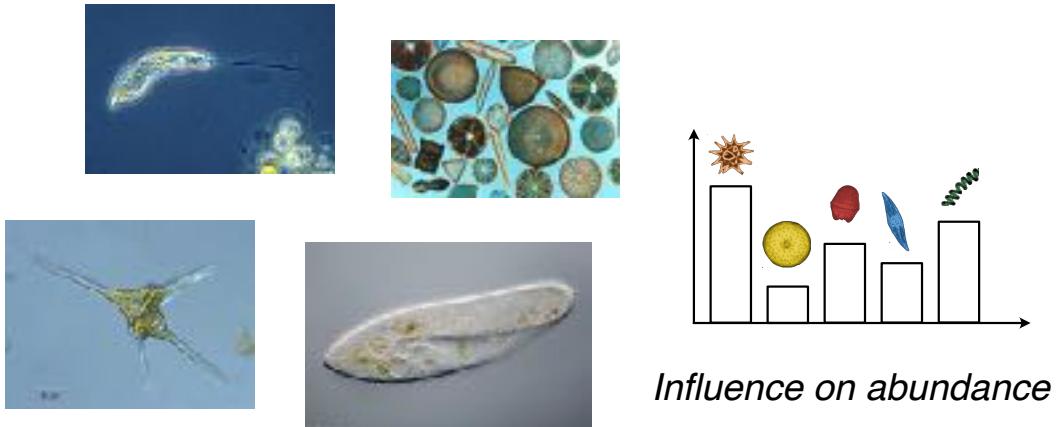
## Classical marker genes - an overview



### Various copy numbers between taxa



*To meet the huge biosynthetic demand, eukaryotic cells contains hundreds to thousands of copies of ribosomal genes. (Kobayashi et al., 2011)*



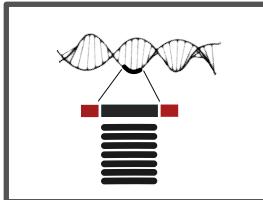
Martin et al., 2022



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# Classical marker genes - an overview

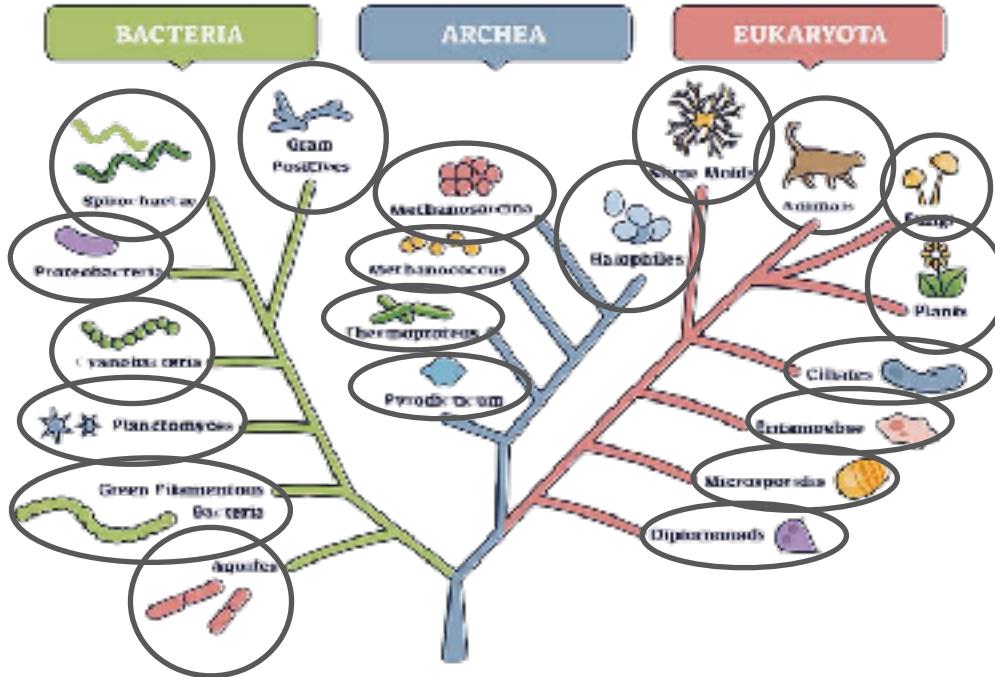


Ribosomal proteins

Nuclear

Mitochondrial

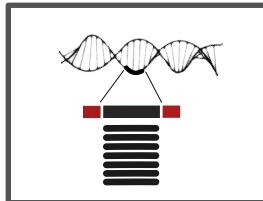
Chloroplast



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# Classical marker genes - an overview

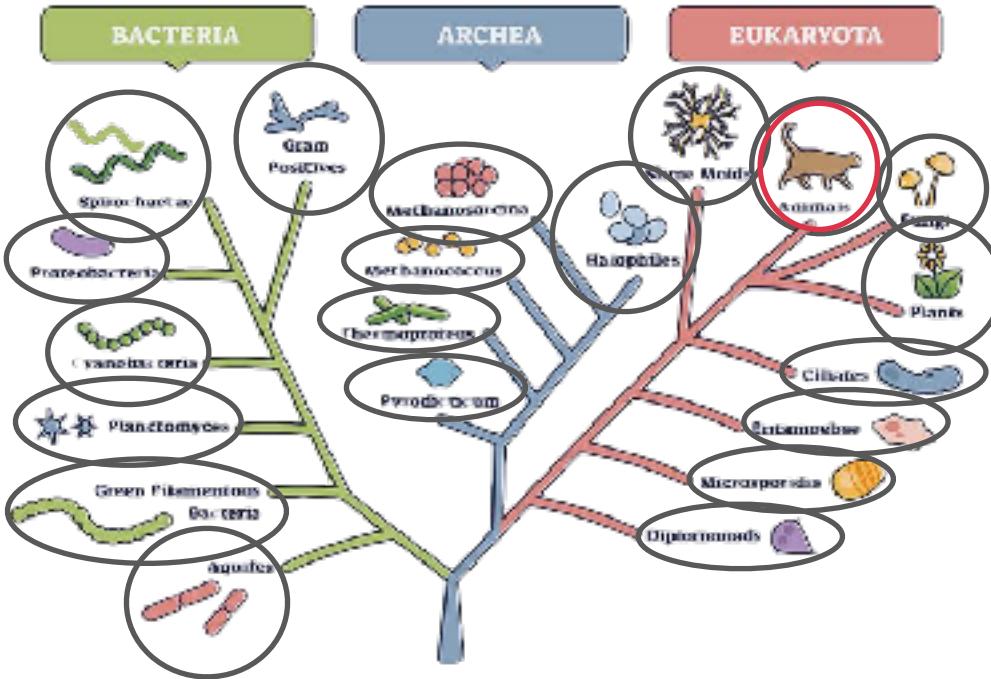


Ribosomal proteins

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Chloroplast



COI

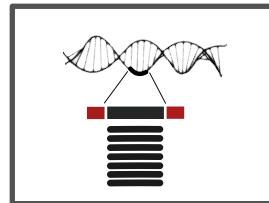
Mitochondrial



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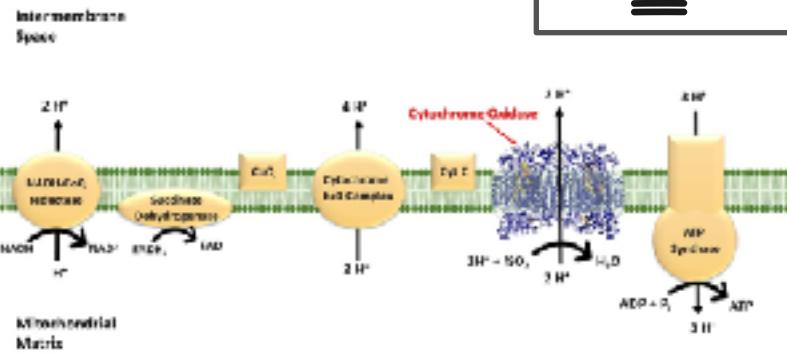
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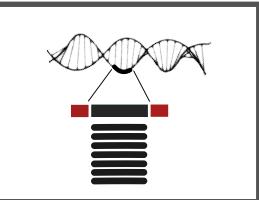
## Classical marker genes - COI gene



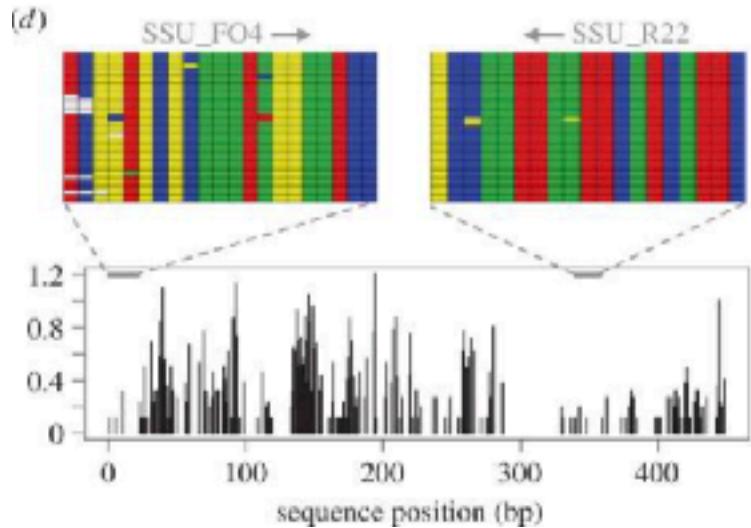
### COI

- COI codes for the Cytochrome Oxidase subunit I  
It is a key mitochondrial enzyme for respiration
- Historically it is used for barcoding of animals. lot of primers has been designed for various animals group
- COI often provides a better taxonomic resolution than nuclear rRNA (18S)
- But contrary to ribosomal genes, it is complex to define universal primers.





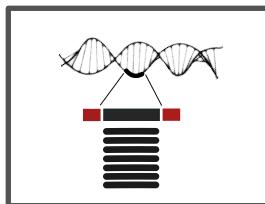
## Nuclear 18S



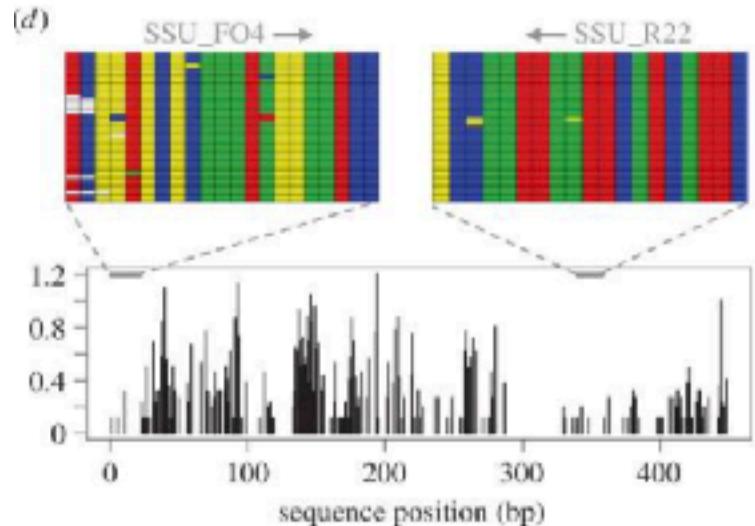
*From more than 40 species of insects among 25 different orders (Deagle et al. 2014)*



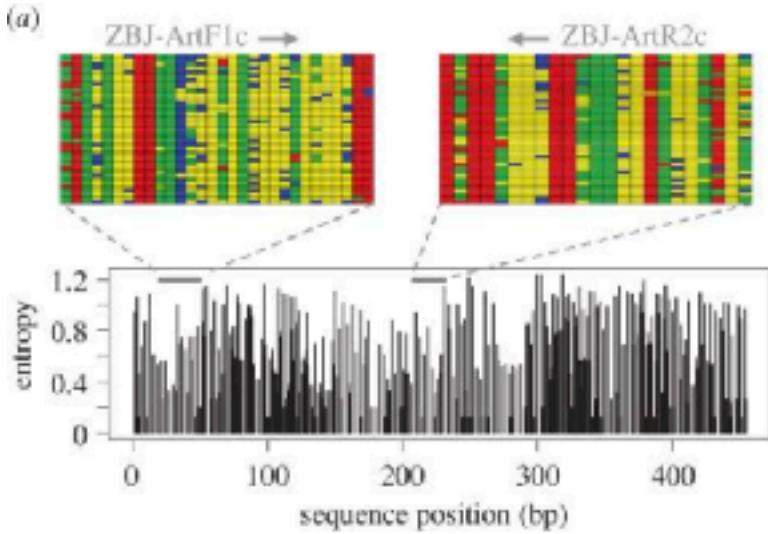
## Classical marker genes - COI gene



### Nuclear 18S



### COI



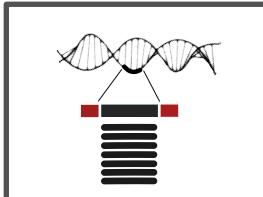
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# Classical marker genes - an overview

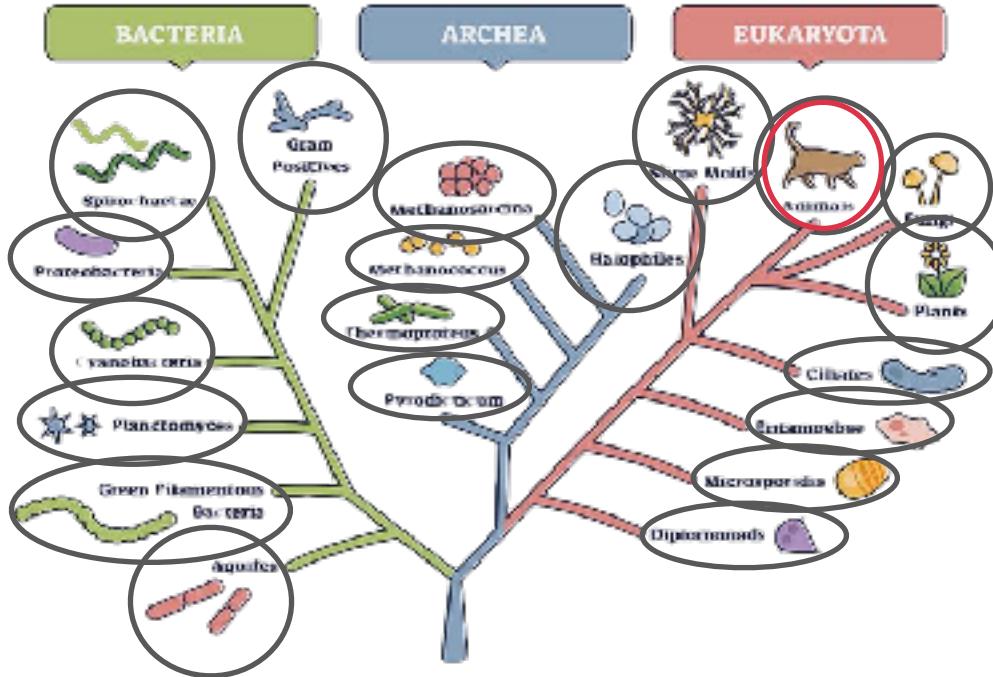


Ribosomal proteins

Nuclear

Mitochondrial

Chloroplast



COI

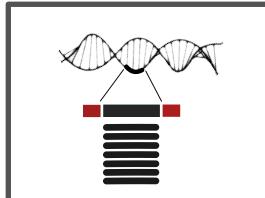
Mitochondrial



Funded by European Union

[www.biolaweb.com](http://www.biolaweb.com)

# Classical marker genes - an overview

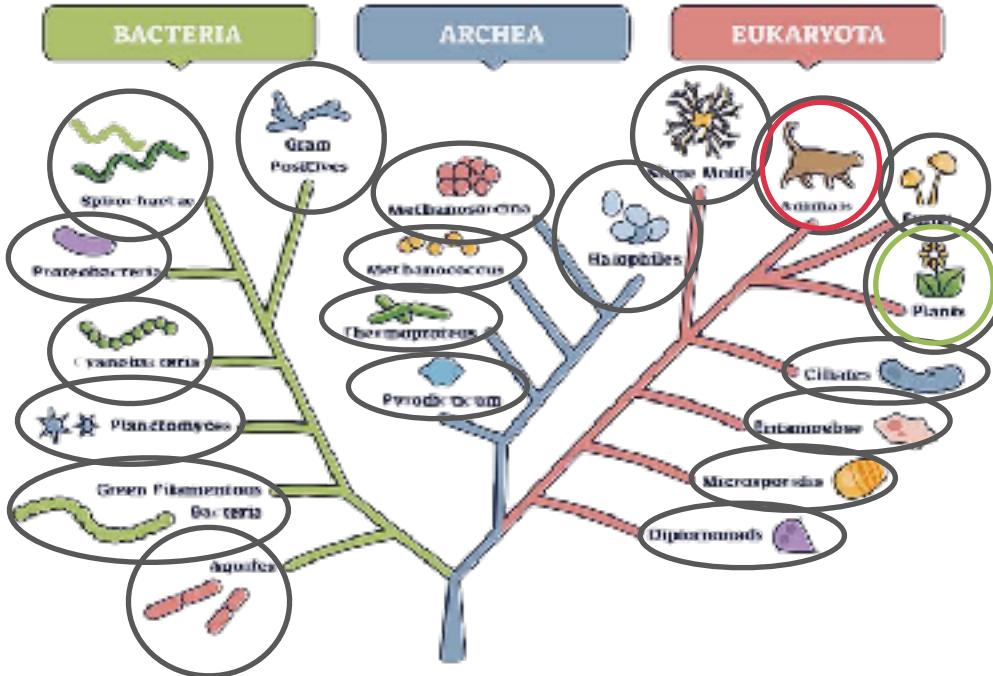


Ribosomal proteins

Nuclear

Mitochondrial

Chloroplast



COI

Mitochondrial

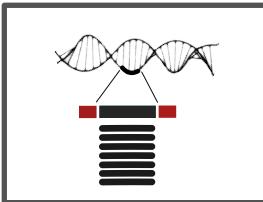
RbcL/matK

Chloroplast





## Classical marker genes



### RbcL/MatK - chloroplastic genes to target photosynthetic organisms

- For plant, low substitution rates of mitochondrial DNA - COI is not a good target
- Genes present in chloroplasts - target specifically photosynthetic organisms

MatK : plastid genes that codes for an intron maturase

RbcL : codes for the large subunit of RuBisCo, a key enzyme of photosynthesis.

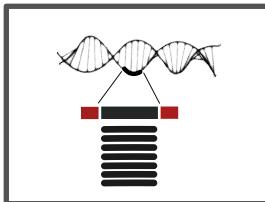
Difficulties to get target all the phytoplanktonic diversity

Ribosomal genes (16S)

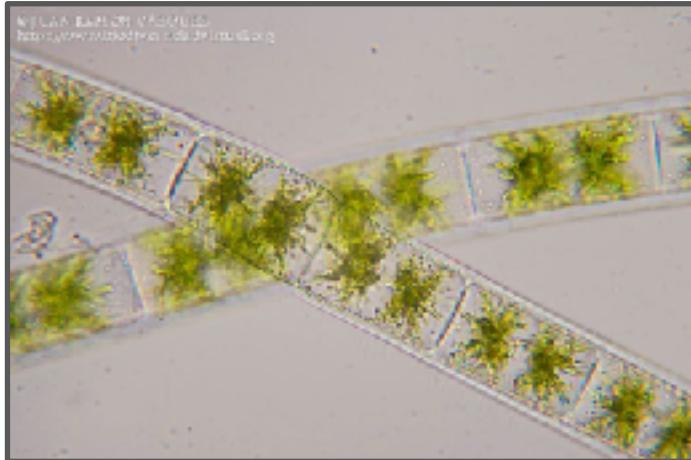




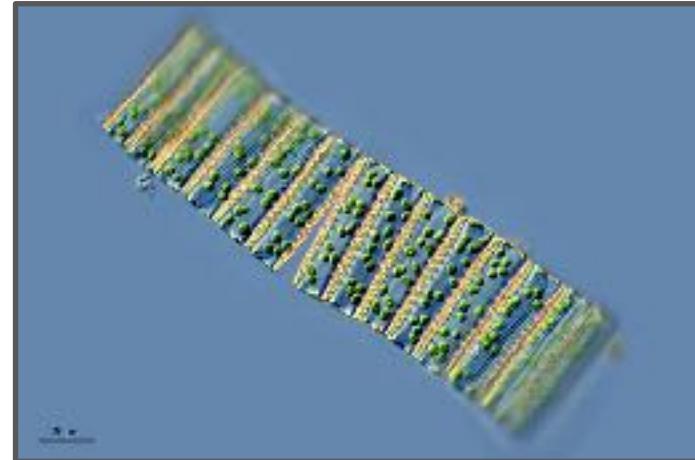
## Classical marker genes - an overview



- A marker gene in chloroplasts and mitochondria can be present in variable numbers in the cell



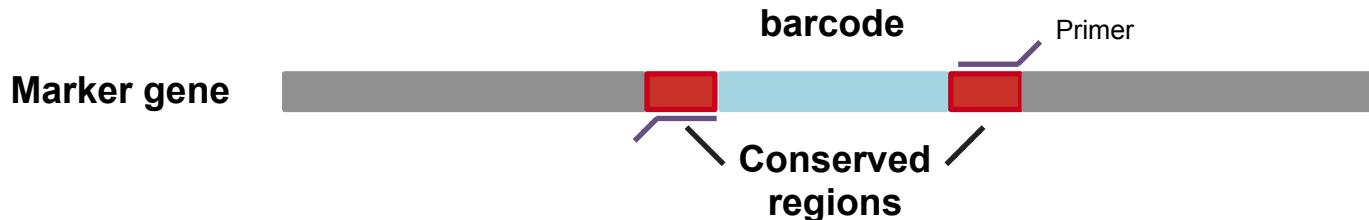
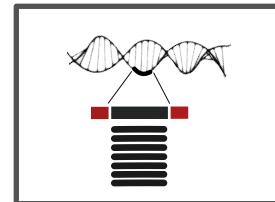
*Zygnema sp.* 1-2 chloroplasts/cell



*Diatoma sp.* > 10 chloroplasts/cell



## Barcode selection - a fundamental criteria



1 Universal

2 Conserved regions

3 Variable enough

4 Match sequencing technology size

5 Represented in reference libraries

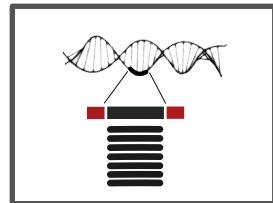
How primers are designed to hybridize only in the DNA of all targeted organisms?

### Specificity



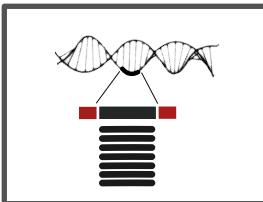
Funded by European Union

[www.biolaweb.com](http://www.biolaweb.com)



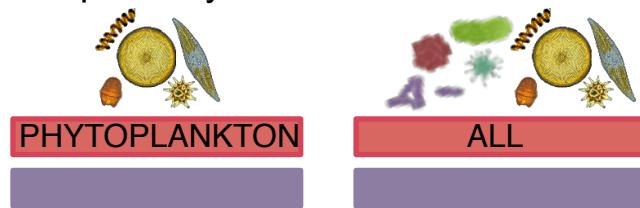
## Conserved regions : specificity

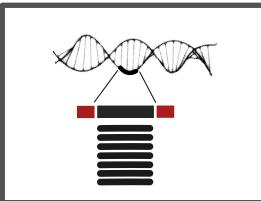




## Conserved regions : specificity

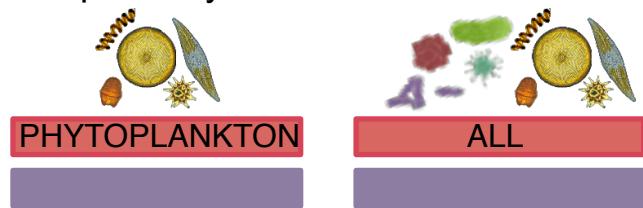
Primer specificity :





## Conserved regions : specificity

Primer specificity :



Primer degenerescence :

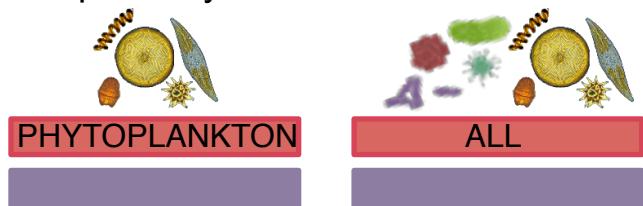
A sequence alignment of four primers against a target DNA template. The target sequence is GAGCCTAATAACC. The first primer (yellow) matches perfectly. The second primer (orange) has a mismatch at position 4 (T instead of C). The third primer (red) has a mismatch at position 4 (T instead of C) and a frameshift starting at position 6. The fourth primer (purple) has a mismatch at position 1 (C instead of G) and a frameshift starting at position 2.

Code	Description
M	AC
R	AG
W	AT
S	CG
Y	CT
K	GT
V	ACG
H	ACT
D	AGT
B	CGT
N	ACGT

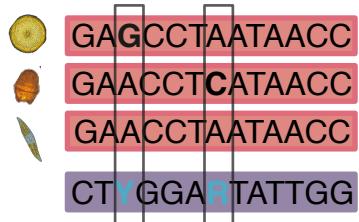


## Conserved regions : specificity

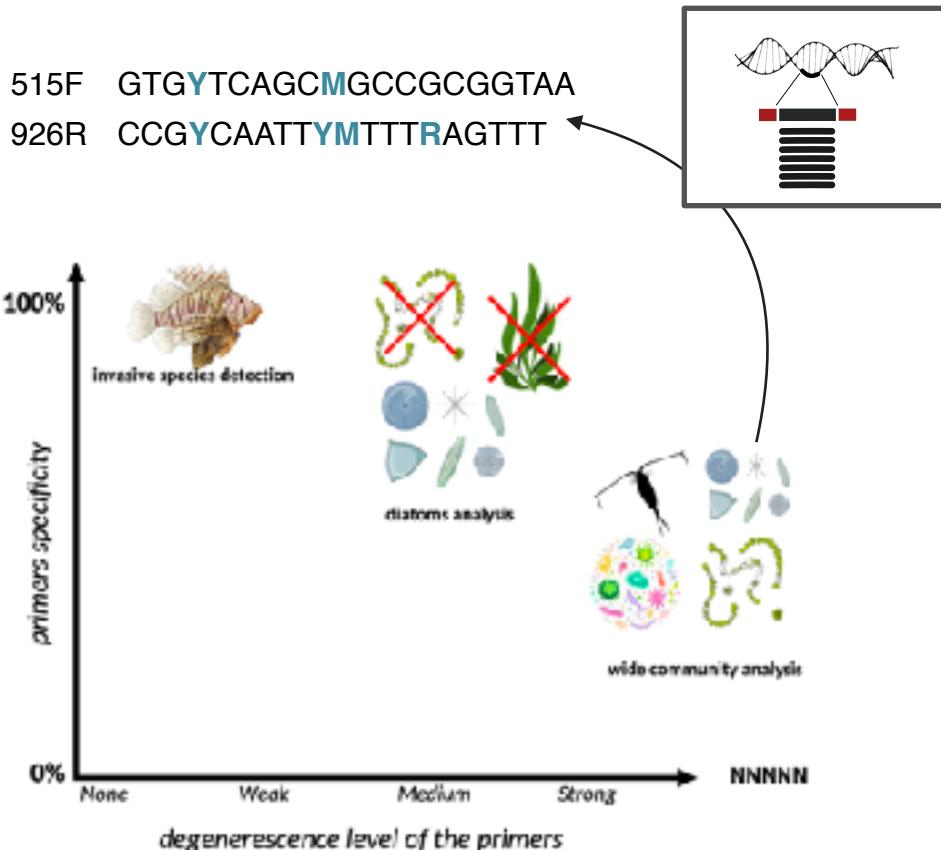
Primer specificity :



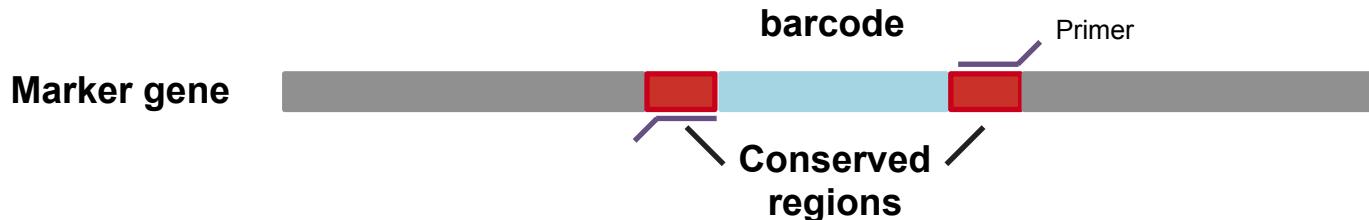
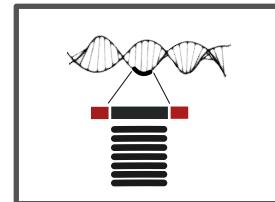
Primer degenerescence :



Code	Description
M	AC
R	AG
W	AT
S	CG
Y	CT
K	GT
V	ACG
H	ACT
D	AGT
B	CGT
N	ACGT



## Barcode selection - a fundamental criteria



1 Universal

2 Conserved regions

3 Variable enough

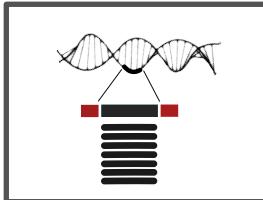
4 Match sequencing technology size

5 Represented in reference libraries

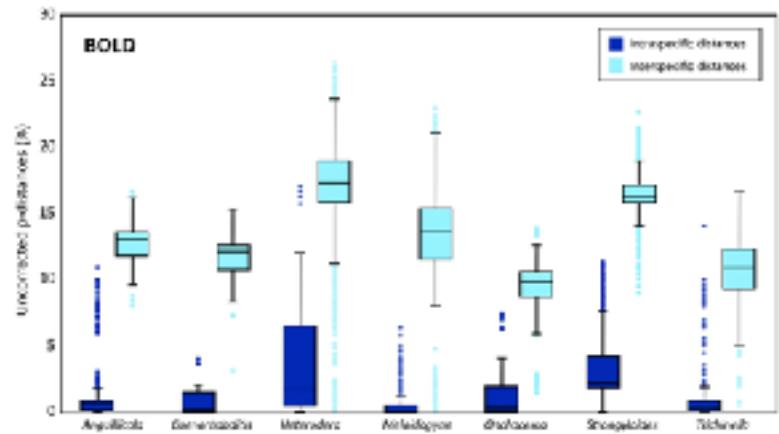
Barcode-gap efficiency  
for a great diversity of  
organisms in  
metabarcoding



## Variable enough

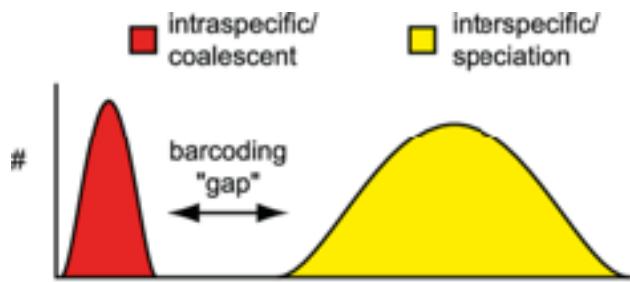


Depends on the taxonomic group



Tresoldi-Gonsaves et al., 2021

COI of various genera of worms  
(Nematoda)



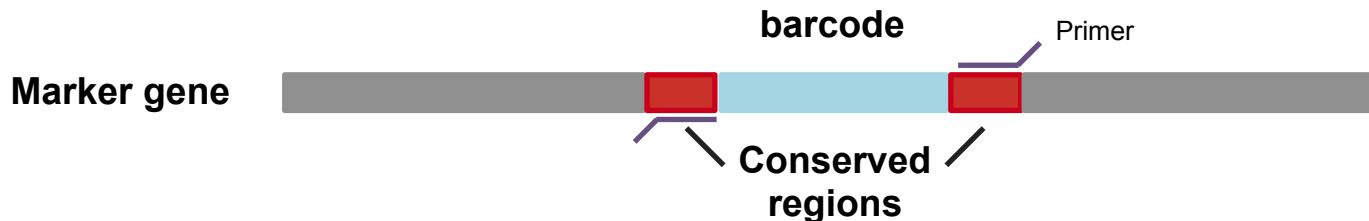
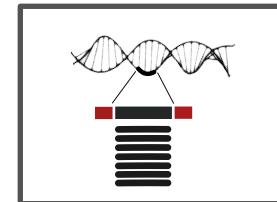
There is often a trade-off between capacity to target a large diversity and the barcode variability



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## Barcode selection - a fundamental criteria



1 Universal

2 Conserved regions

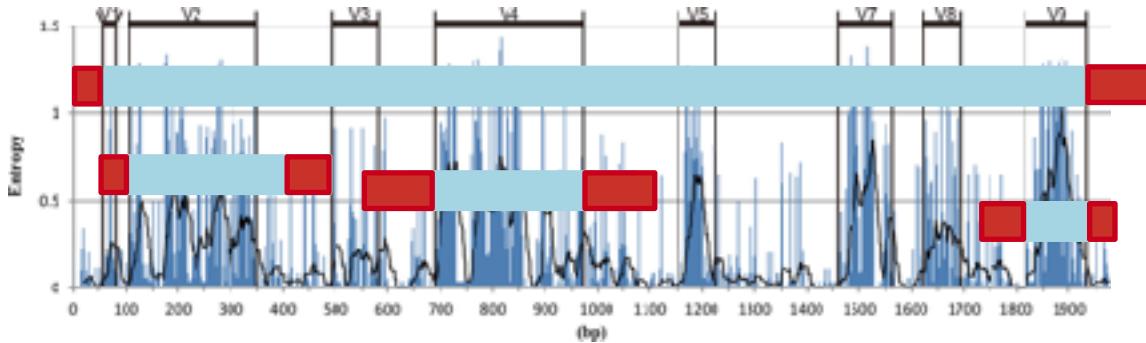
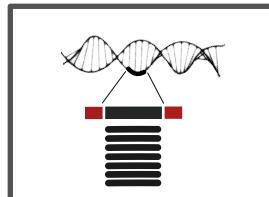
3 Variable enough

4 Match sequencing technology size

5 Represented in reference libraries

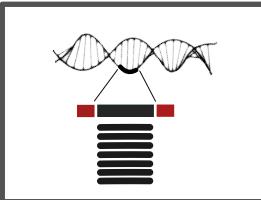


## Match sequencing technology size

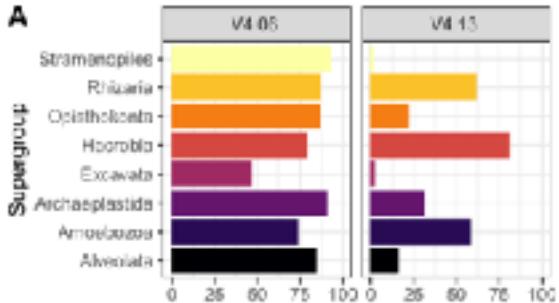
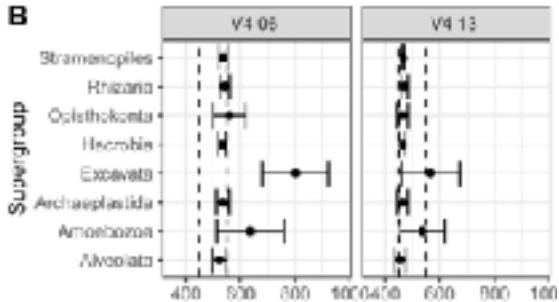


ADNr 18S :	1800bp
ADNr 16S :	1500bp
Rbcl :	1400bp
MatK :	1500bp
COI :	650bp

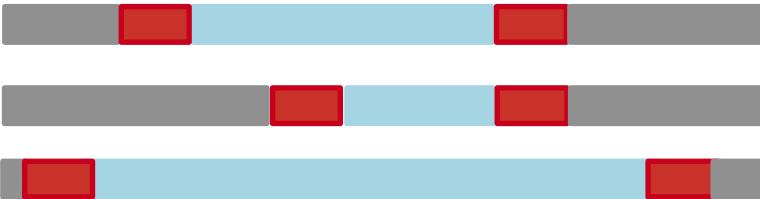




## Match sequencing technology size

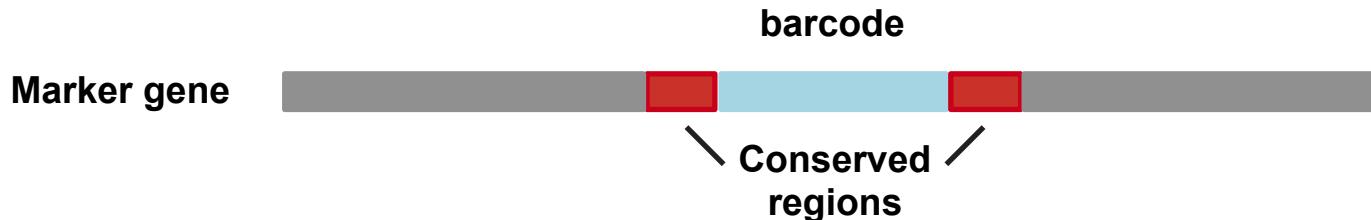
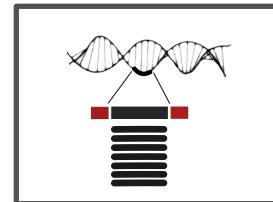
**A**

**B**


For the same primers, the amplicon size will vary between major supergroup of protists



*A broader diversity of protist will be recovered with long-read sequencing*

## Barcode selection



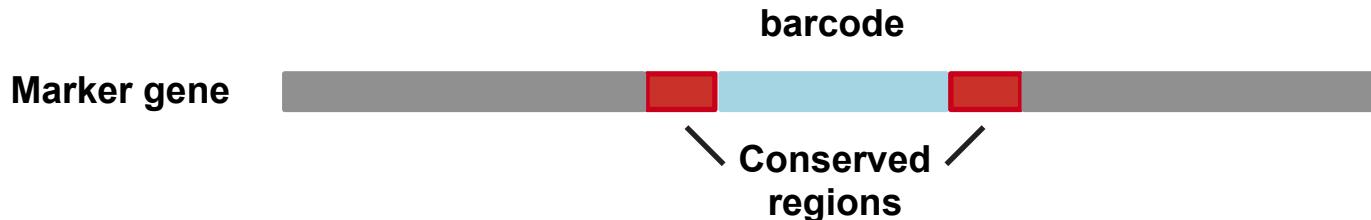
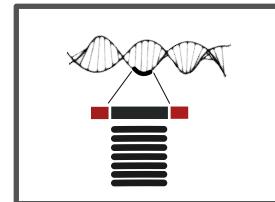
- 1 Universal
- 2 Conserved regions
- 3 Variable enough
- 4 Match sequencing technology size
- 5 Represented in reference libraries

**The perfect barcode does not exist**

*Particularly if you want to target very diverse groups of organisms (e.g. all the protists, all the macroinvertebrates, all the phytoplankton)*



## Barcode selection

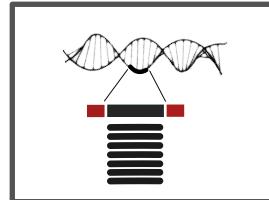


- 1 Universal
  - 2 Conserved regions
  - 3 Variable enough
  - 4 Match sequencing technology size
  - 5 Represented in reference libraries
- The perfect barcode does not exist**

*Find the one that best suits to your research question*



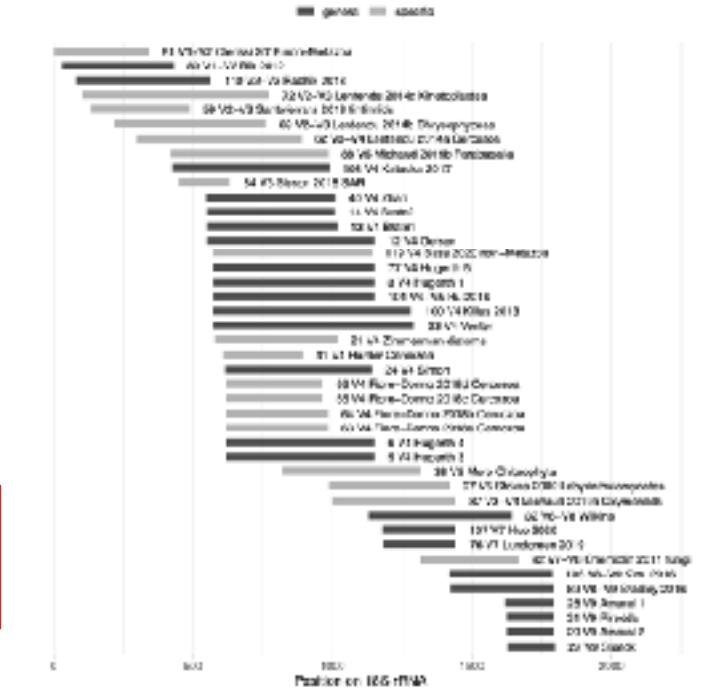
# How to select a barcode ?



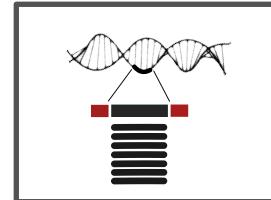
In literature

- Comparison between dataset
- Known bias
- Tested

- Possibly not adapted to your study/environment



# How to select a barcode ?



In literature

- Comparison between dataset
- Known bias
- Tested

- Possibly not adapted to your study/environment



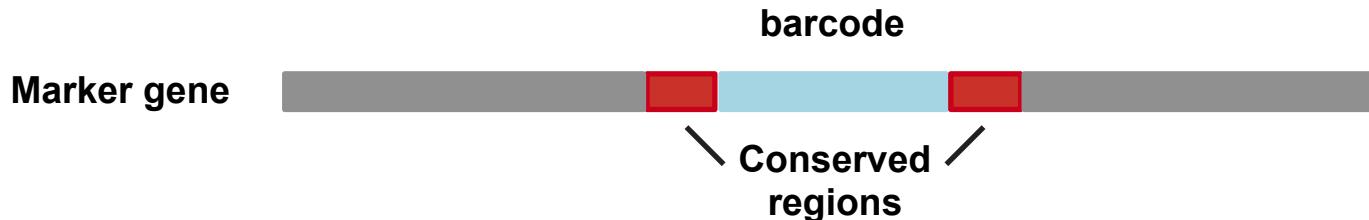
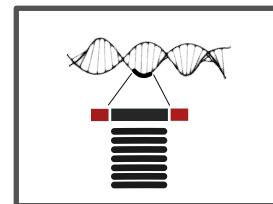
- Most adapted to your study
- Up to date reference libraries

Time-consuming !

- A lot of optimization steps



## Barcode selection



1 Universal

2 Conserved regions

3 Variable enough

4 Match sequencing technology size

5 Represented in reference libraries

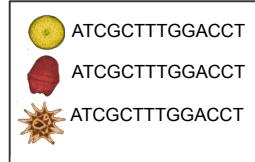
« *The power of DNA metabarcoding is directly proportional to data available in reference databases* »



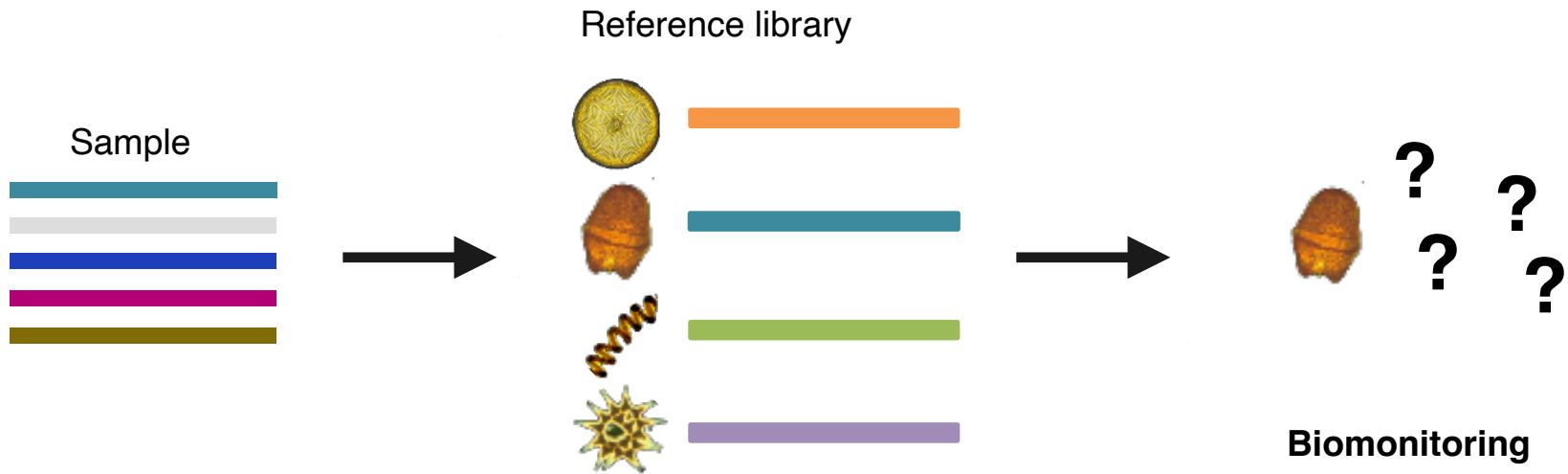
Funded by European Union

[www.biolaweb.com](http://www.biolaweb.com)

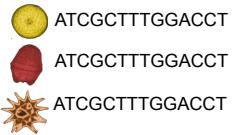
## Reference libraries



*They are a prerequisite to identify your sequence and give it a taxonomy*

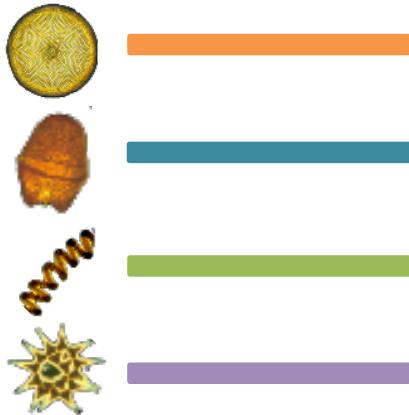


## Reference libraries

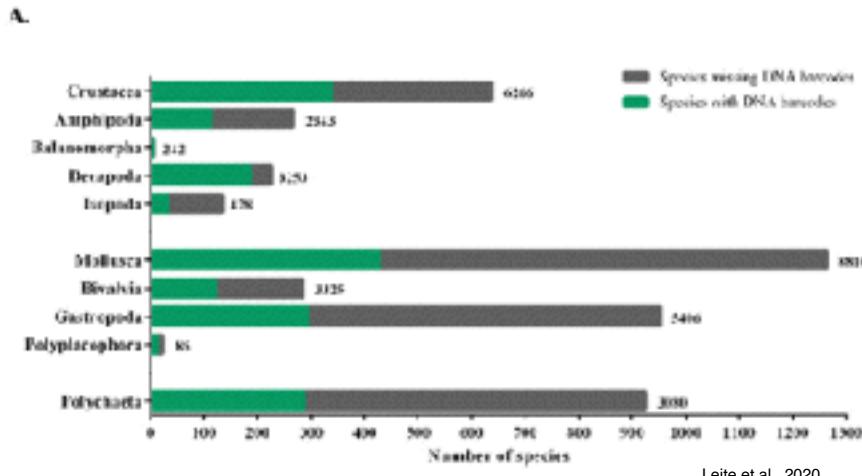


*They are a prerequisite to identify your sequence and give it a taxonomy*

Reference library



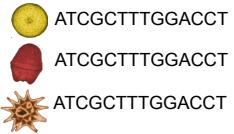
→ The most complete as possible



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[www.biolaweb.com](http://www.biolaweb.com)

## Reference libraries



*They are a prerequisite to identify your sequence and give it a taxonomy*

Reference library



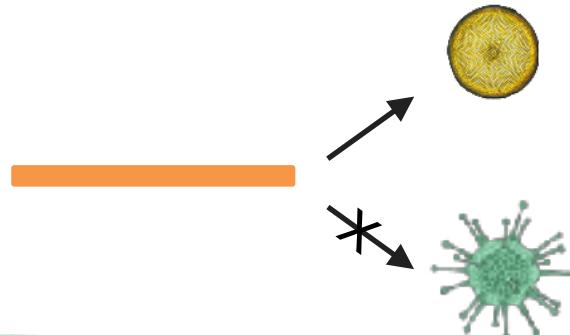
→ The most complete as possible



→ The most curated as possible



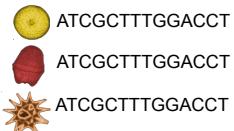
*Up to date taxonomy*



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## Reference libraries - ribosomal genes



### Generalists



#### SILVA SSU 138.1 update release

	SSU Parc	SSU Ref NR 99	LSU Parc	LSU Ref NR 99
Minimal length	300	1200/900	300	1900
Quality filtering	basic	strong	basic	strong
Guide Tree	no	yes	no	yes
Release date	27.06.20	27.06.20	27.06.20	27.06.20
Aligned rRNA sequences	9,469,124	510,508	1,312,534	95,286

For bacteria, archaea and eukaryotes  
Both nuclear and plastid SSU and LSU

### Specialists



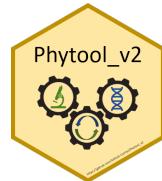
Nuclear  
18S

200,000  
sequences

PhytoRef

Plastidal  
16S

6486  
sequences



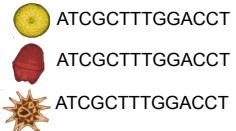
Nuclear 18S  
Plastidal  
16S, 23S

18S : 16654  
16S : 8479  
23S : 1997

For protists

For phytoplankton

## Reference libraries - other genes



### Generalists

## BOLD SYSTEMS

**BOLD: The Barcode of Life Data System**  
[www.barcodinglife.org](http://www.barcodinglife.org)

SUJEEVAN RATNASHAM and PAUL D. N. HEBERT

Around 1.3M of COI barcodes



### Specialists

#### Diatbarcode



rbcl

4000-5000  
sequences

Diatoms

#### COIns



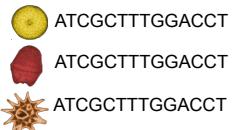
COI

532 000  
sequences

Insects



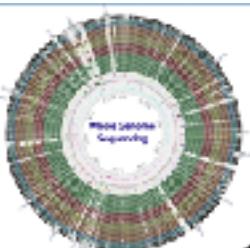
## Reference libraries - non curated database



Your study



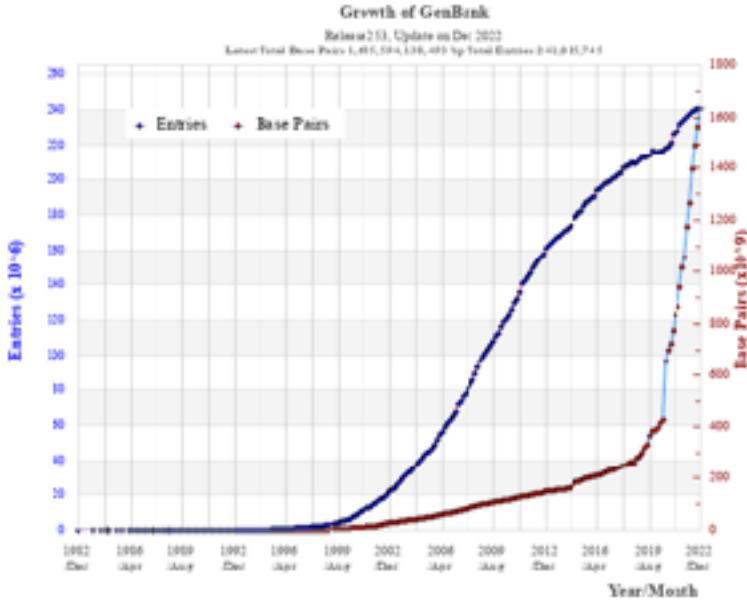
Big eDNA projects



**ENA**  
European Nucleotide Archive

**DDBJ**  
DNA Data Bank of Japan

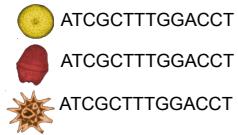
**NCBI**



For GenBank : 240 millions of sequences

High potential but **not curated !**

## Reference libraries



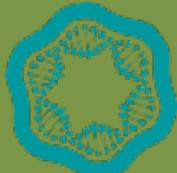
All sequences



Quality-checked



Curated taxonomy



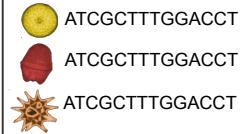
PhytoRef



Funded by European Union

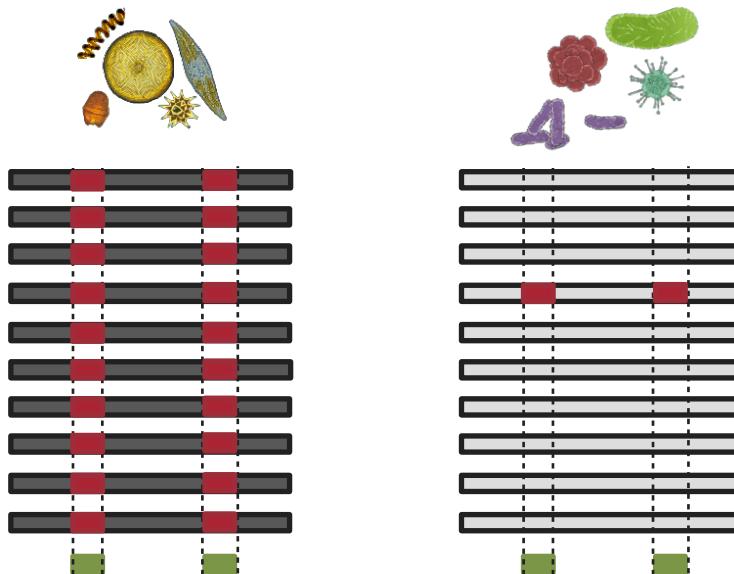
[www.biolaweb.com](http://www.biolaweb.com)

## Reference libraries



But also they are essential to find the best design for the primers

Optimal design of the  
primers



Test for primer specificity



Funded by European Union

[www.biolaweb.com](http://www.biolaweb.com)

## Reference libraries



But also they are essential to find the best design for the primers

Optimal design of  
primers

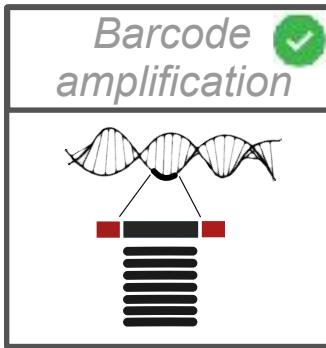
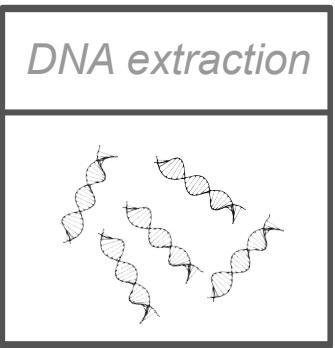
st for primer specificity



The screenshot shows the BioPrimers web interface. At the top, there are links for Home, SILVaNGs, Browser, Search (which is highlighted in orange), ACT, Download, Documentation, Projects, Jobs, and Contact. Below this is a search form titled "BioPrimers 2.6". It has two input fields: "Forward Primer (5'...7')" and "Reverse Primer (7'...5')". Under "Database", it says "SILVA REFERENCE SSU 128L". Under "Mismatches", it says "MAXIMUM NUMBER OF MISMATCHES: 0 mismatches". Under "Sequence Collection", it says "RefNR". Under "Length of 0 mismatch ends at 3' end", there is a dropdown menu set to "selected". On the right side of the search form, there is a "ProbeDatabase Search" section with a text input field for "Please enter the probe name or sequence" and a note: "This service is provided by probebase.org". Below the search form are two diagrams showing primer designs. The first diagram shows a black horizontal bar with two red boxes (representing primers) and two green boxes below it (representing sequencing). The second diagram shows a similar setup but with a longer black bar and two green boxes below it.



## Metabarcoding steps



*Bioinformatic analysis*



ATCGCTTGGACCT  
ATCGAATTGGAACA  
ATCGCTTGGACCT  
ATCGAATTGGAACA  
ATCGCTTGGACCT  
ATCGAATTGGAACA

