



# Diatom metabarcoding for biomonitoring : 4th part

# F. Rimet, A. Bouchez

Application of diatom biomonitoring, Source of bias, Intercalibration exercise



# Schedule

# 1-Application of diatom biomonitoring to rivers



- Imitation: use of existing diatom indices
- Index based on phylogenetic inference of ecological profiles
- Taxonomy free indices
- Which bootstrap % to use for taxonomic assignation?
- 2- Source of bias
- 3- Intercalibration exercise



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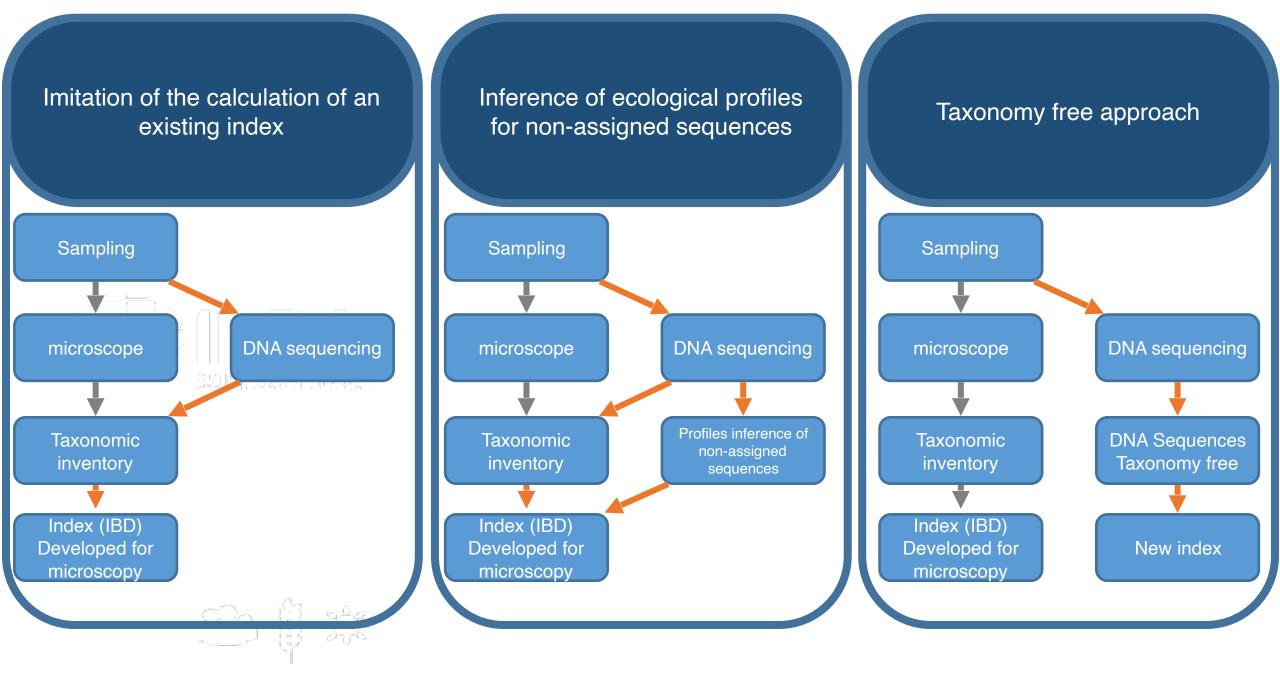
# Schedule

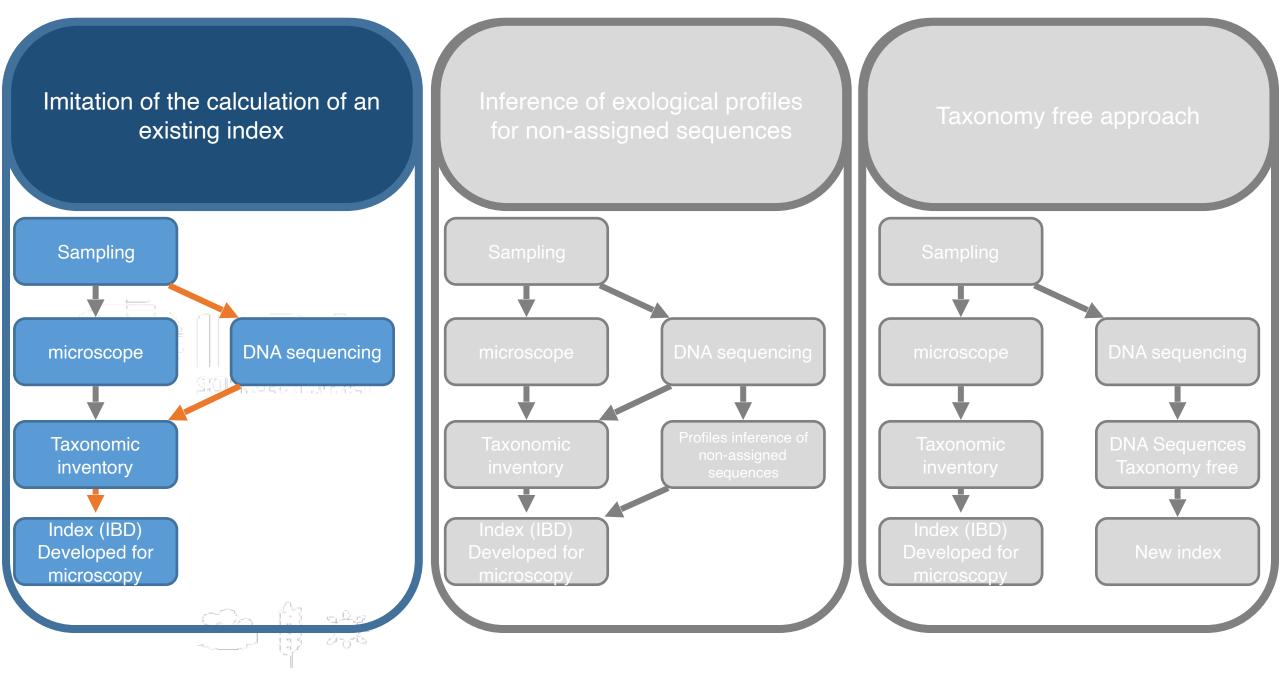
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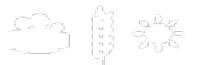
#### Floristic list



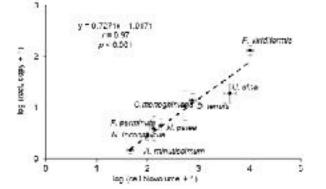
Floristic list modified



Taxon	Séquences %	biovol (µm3)	CF v2		expressed in %	frustule %
Achnanthidium minutissimum	14,0	76,0	2,2	6,4	68,3	75,0
Amphora pediculus	2,0	72,0	2,1	0,9	10,0	13,0
Navicula cryptotenella	10,0	386,0	5,6	1.8	18,9	10,0
Melosira varians	73,0	14515.0	282,9	0.3	2.7	2,0

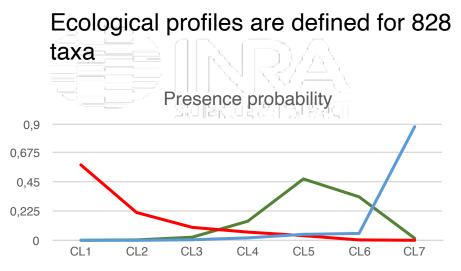


CFV2 = 10^(0,0703\*((LOG(biovolume in μm3))^2,4908))



Comparison of water quality assessment with microscopy and DNA based inventories > Application to France and the IBD

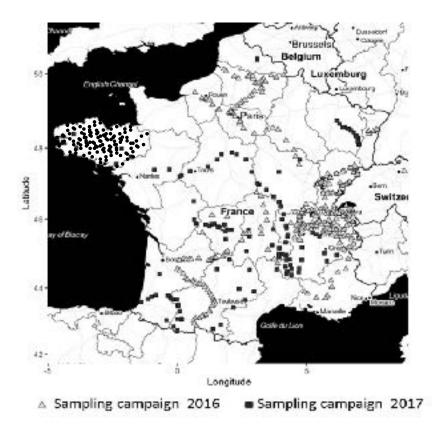
IBD : « Indice Biologique Diatomées » French diatom index used in routine to assess river quality for the WFD



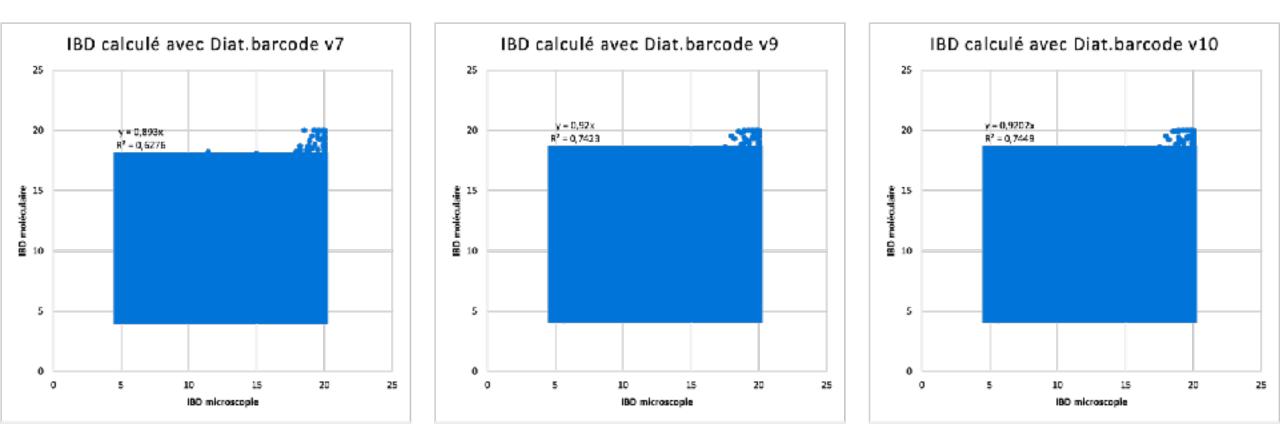
Achnanthidium pyrenaicum (Hustedt) Kobayasi Nitzschia capitellata Hustedt in A, Schmidt et al, var, capitellata Navicula cryptotenella Lange-Bertalot var, cryptotenella



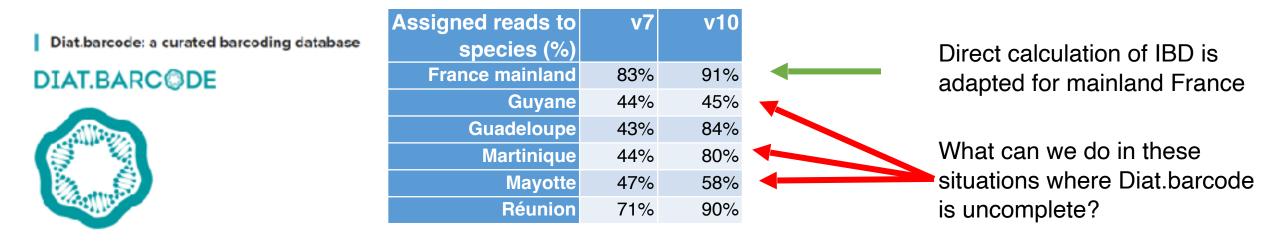
Test carried out on sampling campains 2016, 2017, 2019, in 658 sites



Comparison of water quality assessment with microscopy and DNA based inventories > Application to France and the IBD

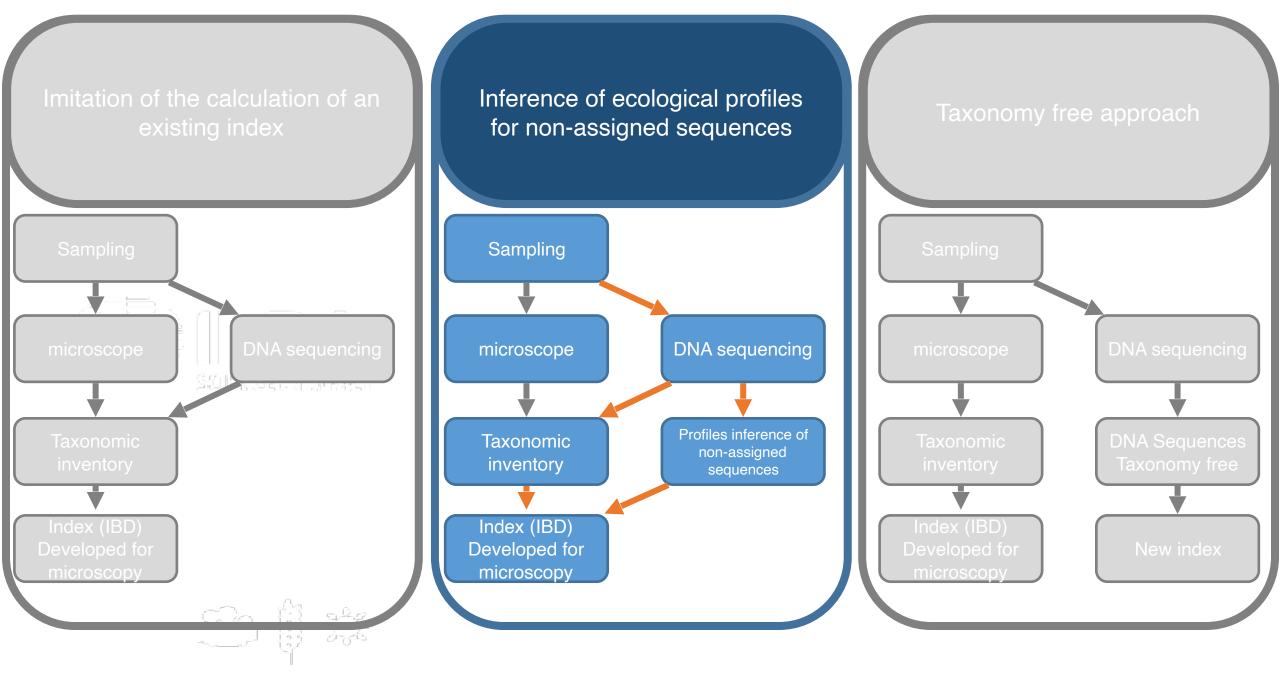


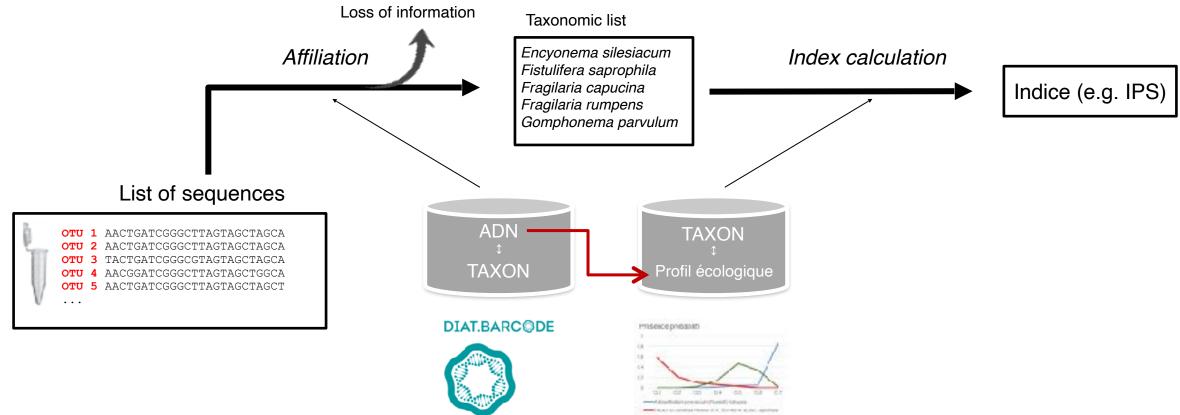
Improvement of the correlation thanks to Diat.barcode completion Very good correlation in mainland France, slope is almost 1:1 What is the proportion of reads assigned to species level depending on Diat.barcode versions (v7 and v10) for 6 different regions?



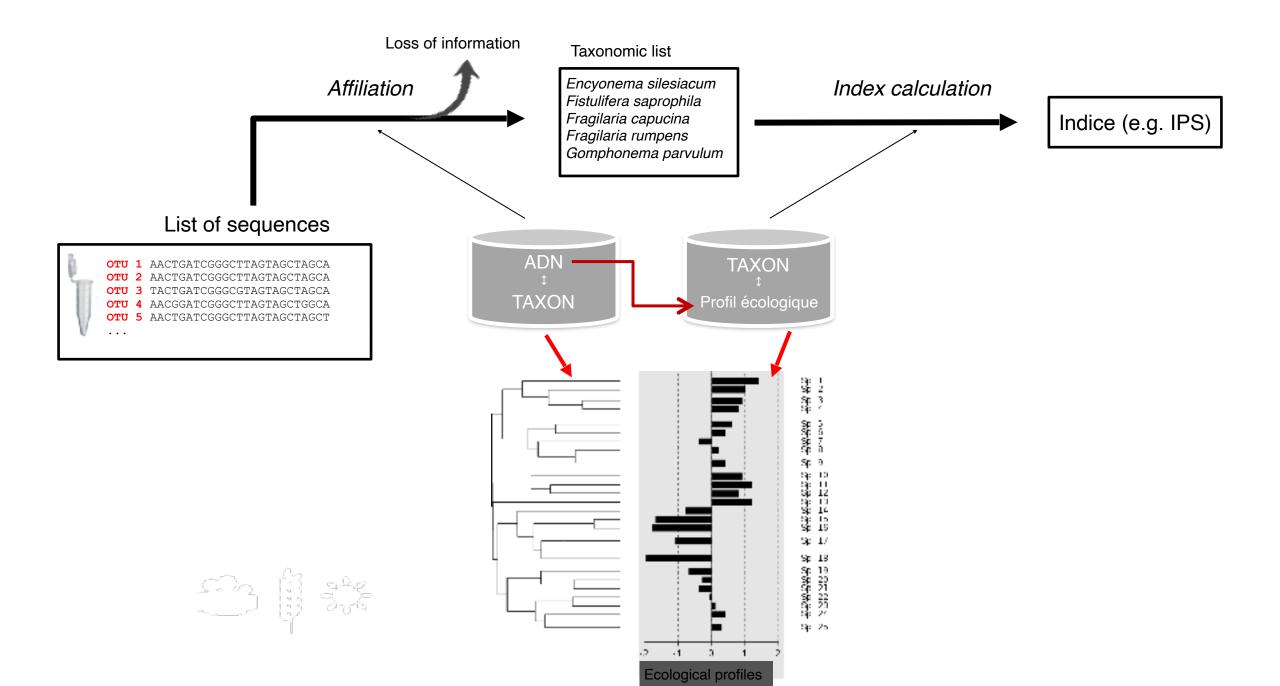
# To sum up:

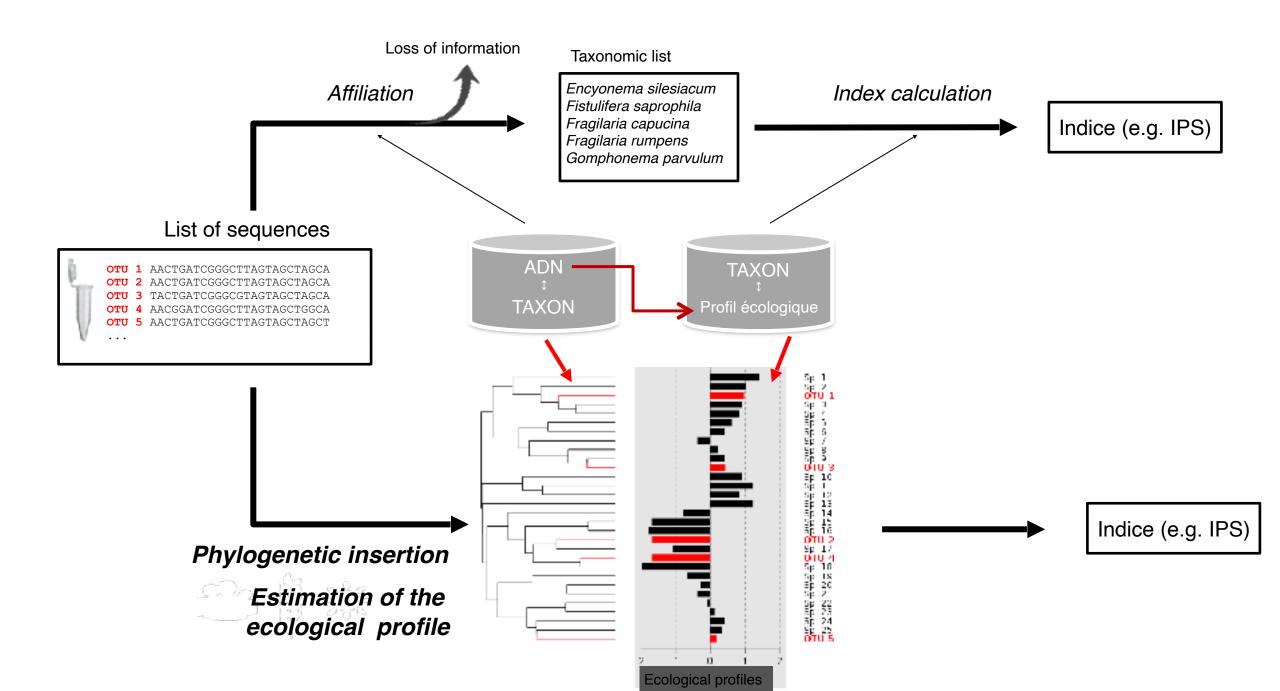
Method adapted when the reference library is almost complete: Microscopy and DNA are very well correlated. However, a few differences (eg. Fragile frustules, dead frustules, cryptic and species difficult to identify).





유할옷

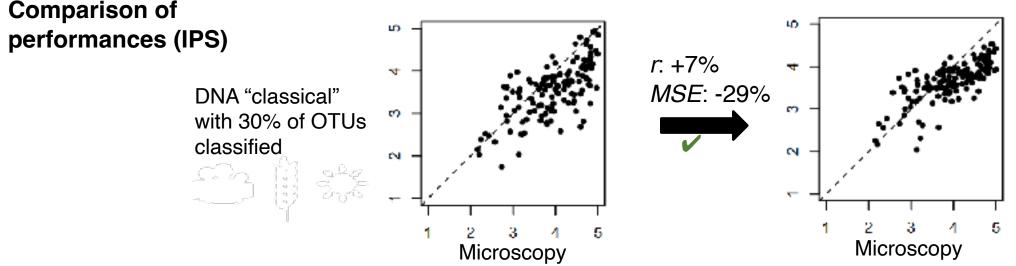




There is a phylogenetic signal of the ecological profile of diatoms (Keck et al. 2016 J. Appl Ecol)

# In situ test (139 sites, mainland France) (Keck et al. 2018, Mol Ecol Res)





DNA "classical" 30% of classified OTUs

70% OTUs non-classified but with a profile inferred from the phylogeny

# To sum up

The method:

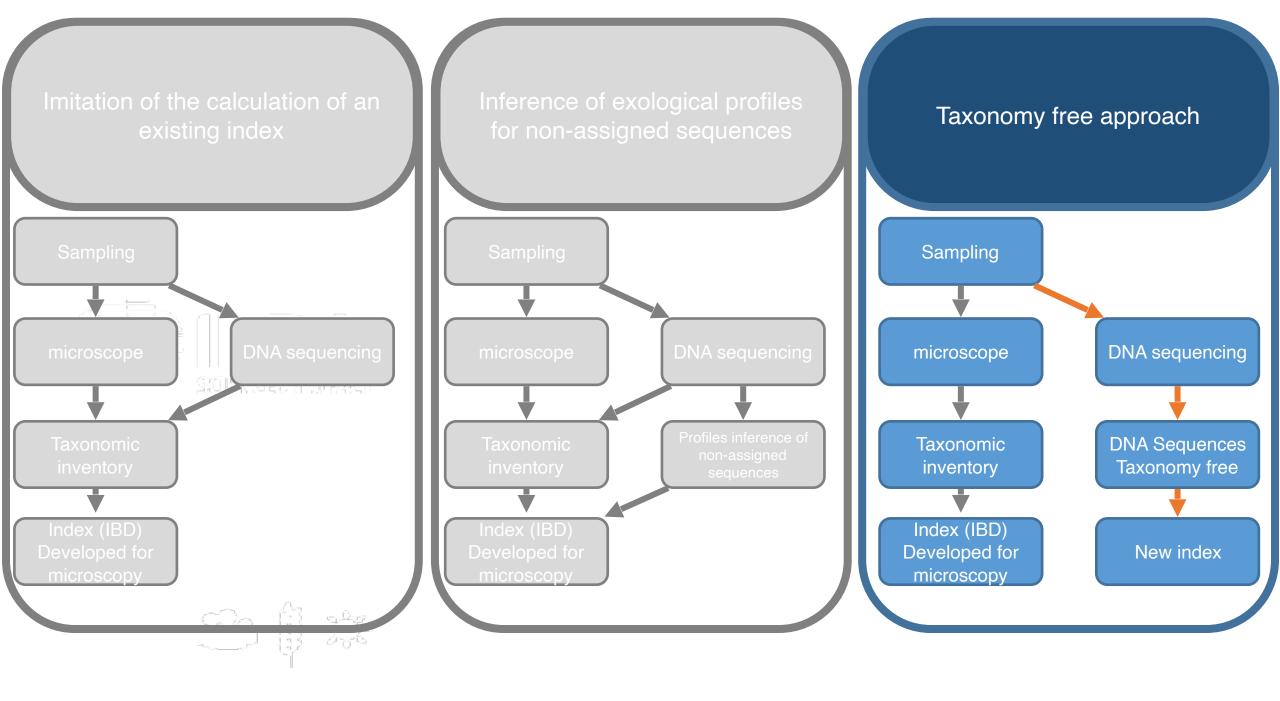
- takes advantage of 100% of the DNA data
- improves the estimation of diatom indices compared to the "classical" approach
- is based on a solid theoretical foundation

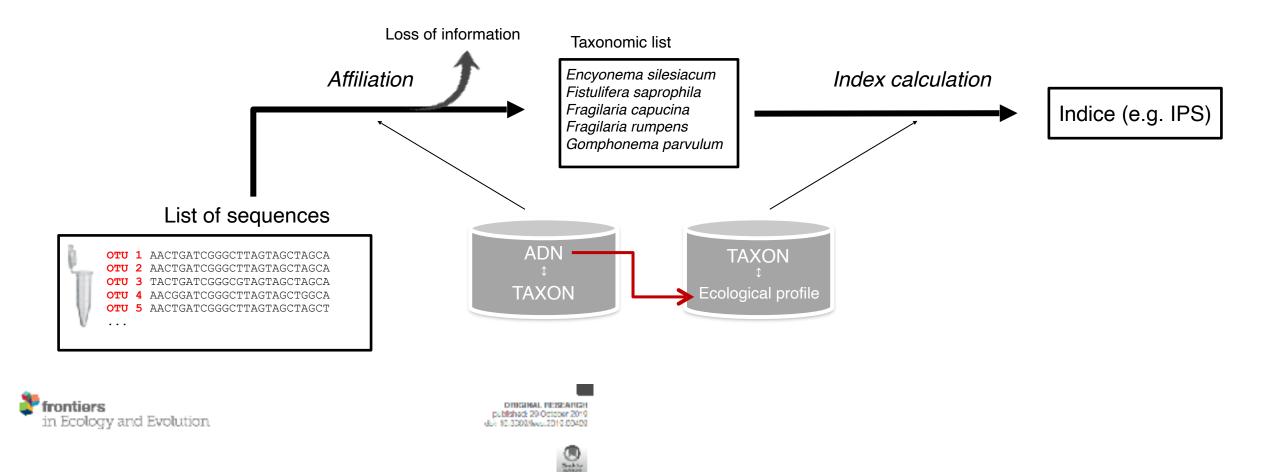
(The phylogenetic signal is a direct consequence of the Darwinian principle of descent with modification)

- is built on existing databases/indices

We can consider this method as a transition to de-novo molecular indices such as:

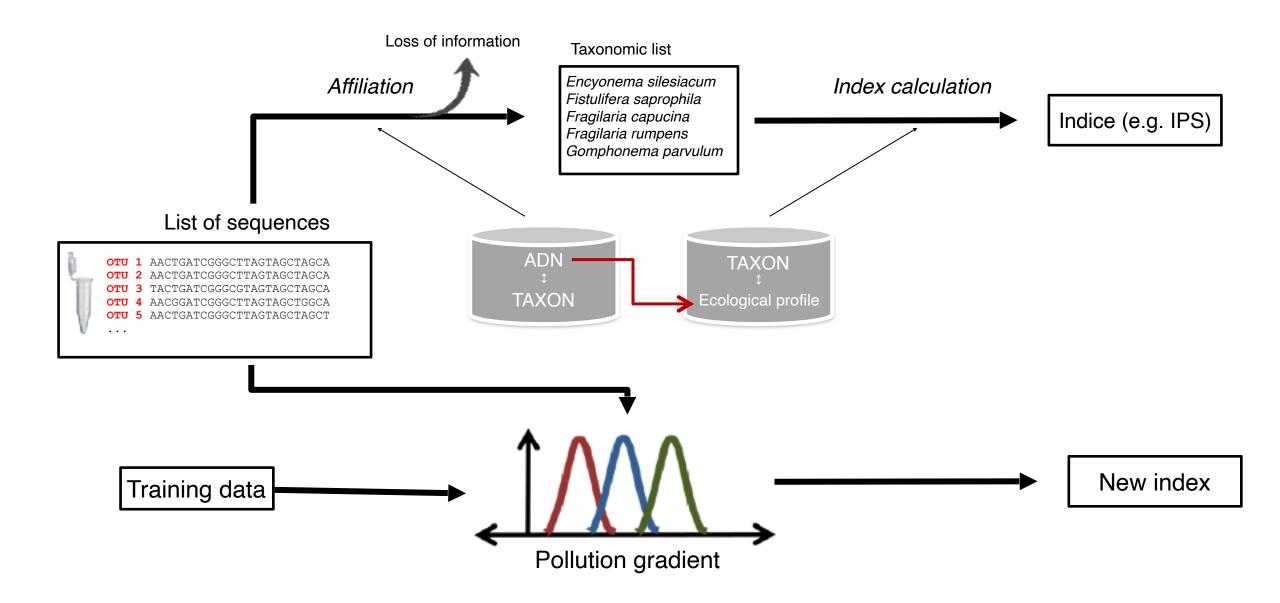
- Taxonomy-free indices
- Supervised machine learning indices



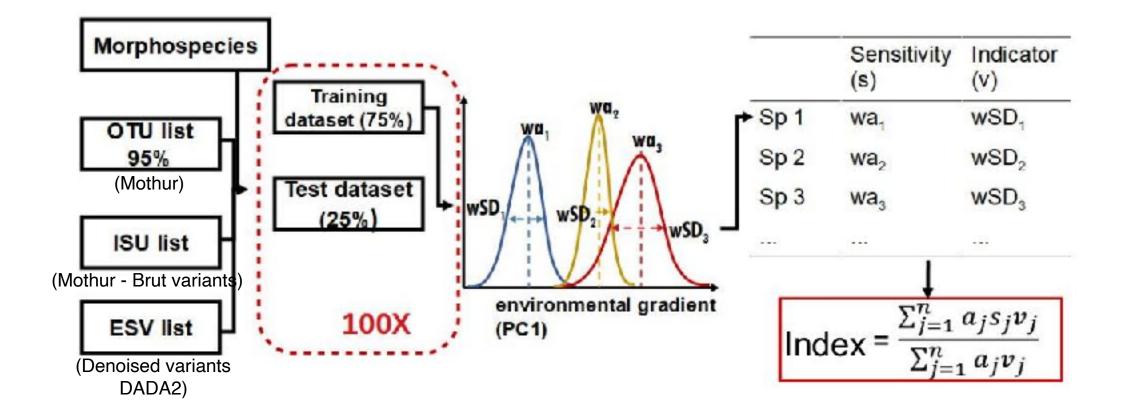


Diatom DNA Metabarcoding for Biomonitoring: Strategies to Avoid Major Taxonomical and Bioinformatical Biases Limiting Molecular Indices Capacities

Kôlmán Tapolozal <sup>134</sup>, François Keck<sup>2,4</sup>, Agnés Bouchez<sup>4,5</sup>, Frédèric Rimet<sup>4,5</sup>, Maria Kahlari<sup>5</sup> and Valentin Vesseion<sup>44</sup>



#### Strategy for the development and test of indices:



ISU: to keep good quality DNA reads using the <u>trim.seqs()</u> command and the following parameters: a sequence length of  $263 \pm 10$  bp (rbcL barcode length without primers), a <u>Phred</u> quality score  $\geq 23$  over a moving window of 25 bp, 0 ambiguities ("N"), a maximum <u>homopolymer</u> length of 8 bp

OTU: (i) ISUs were aligned using the <u>align.seqs()</u> command and poorly aligned reads were removed using the command screen.seqs(start=28, optimize=end, criteria=90); (ii) we used the <u>pre.cluster()</u> command to denoise sequencing errors by preclustering rare ISUs with related more abundant ones (1 bp threshold); (iii) detection of <u>chimeras</u> was performed using the chimera.vsearch() command; Raw sequences

Length / quality filter

Dereplication into ISU

Chimera removal

Taxonomic assignation of ISU (ex: 60, 70, 80%)

Clustering into OTU (ex: Furthest neighbor, Opticlust) (ex: 95%)

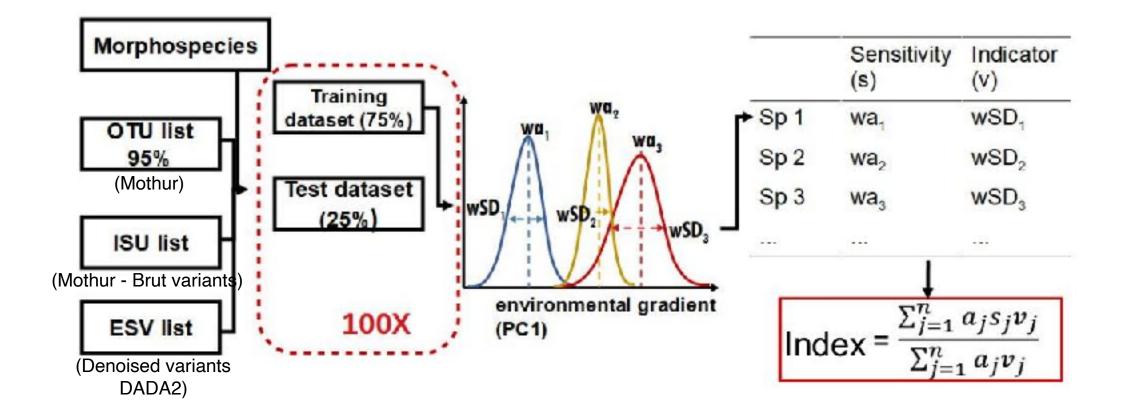
Taxonomic assignation of OTU (ex: threshold 60, 70, 80%)

Molecular inventory



MOTHUR

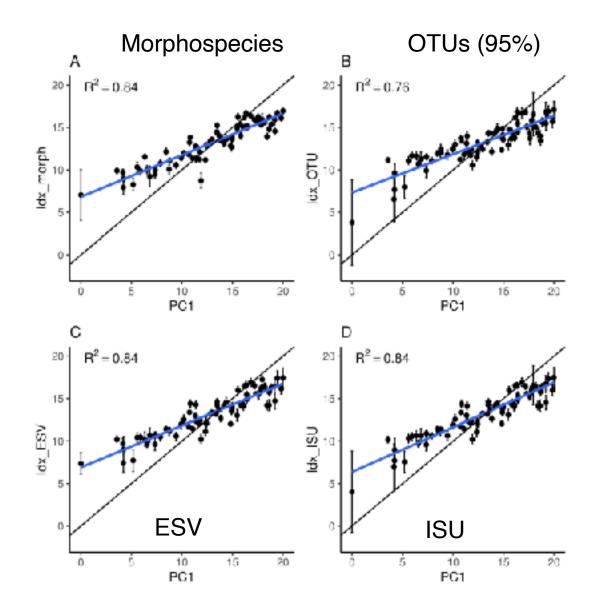
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#### Results



- The 4 indices are working quite well
- Morpho, ISU, ESV indices are very similar
- The OTU index is the least good

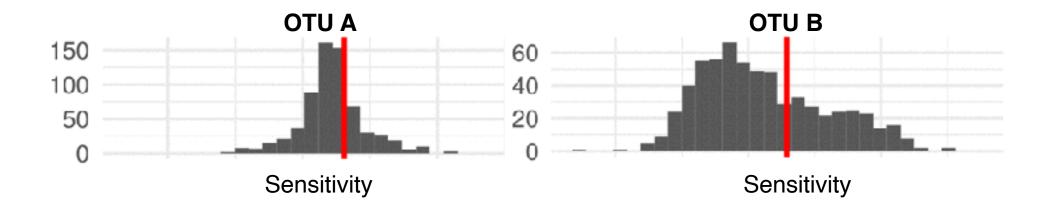
#### Why does the ISU (non-denoised) index work?

- Errors probably removed with the quality filtering
- Errors have probably an ecological profile similar to their original sequence

This questions the need for denoising for bioindication purpose

#### Why does the OTU index work less well?

OTUs group sequences based on genetic similarity and not on ecology



Optimum OTU = optimum of variants

Optimum OTU ≠ optimum of variants

# To sum up

- Taxonomy-free approaches work well (provided you have a suitable dataset for training)
- OTU clustering can hide important ecological information
- Doing without clustering or even denoising (i.e. keeping bioinfo to a minimum) can facilitate the development and standardization of bioindication methods

# To conclude

Diat.barcode: a curated barcoding database

#### DIAT.BARCODE



Ass	signed reads to	<b>v</b> 7	v10	
e	species (%)			
	France	83%	91%	Use of the classical IBD
	métropolitaine			Adapted to phylogenetic assig
	Guyane	44%	45%	of ecological profiles
	Guadeloupe	43%	84% <	
	Martinique	44%	80%	Adapted to taxonomy free whe
	Mayotte	47%	58%	taxonomy is poorly known
-	Réunion	71%	90%	
	Reunion	/   %	90%	



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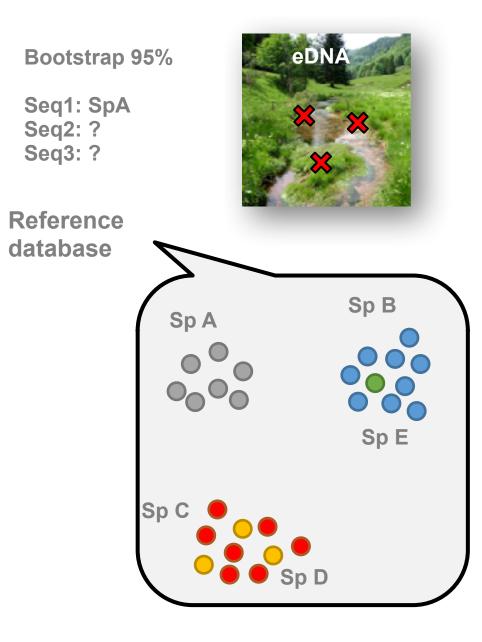
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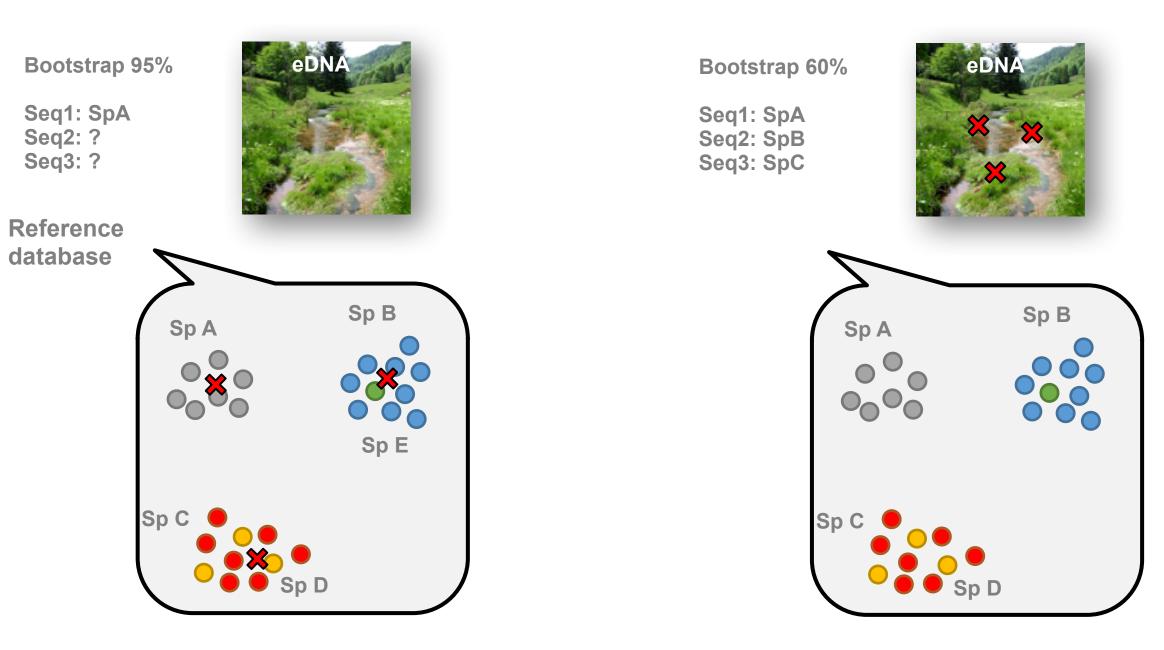


#### What is the bootstrap value for taxonomic assignation?





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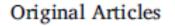
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Diatom metabarcoding applied to large scale monitoring networks: Optimization of bioinformatics strategies using Mothur software

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Raw sequences

Length / quality filter

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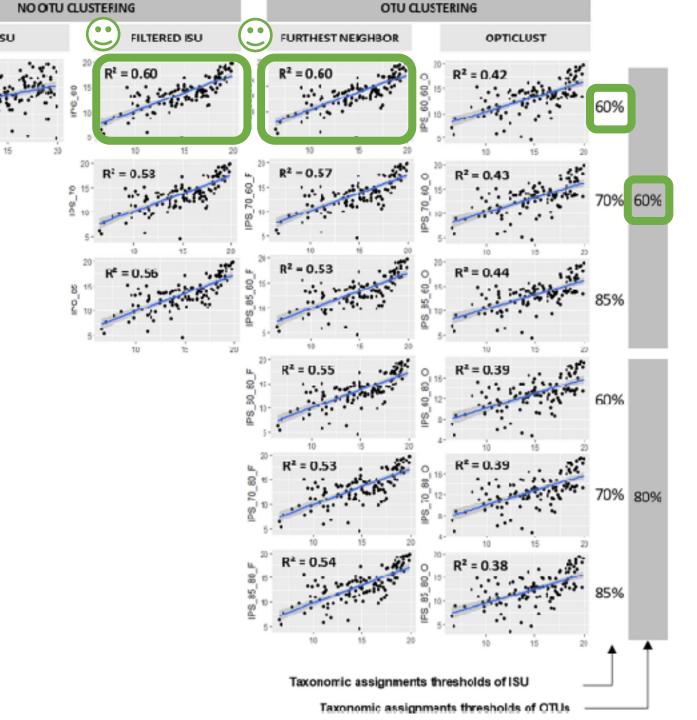
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Correlation between the water quality assessment obtained with microscopy (x axis) and the 16 different bioinformatic strategies (y axis).

The biotic diatom index IPS (indice de Polluosensibilité Spécifique, Cemagref 1982) was calculated. IPS scores vary from 1 (bad quality status) to 20 (good quality status).

IPS scores calculated from ISU with no quality filters (raw data) were poorly correlated to microscopy.

Furthest neighbor and filtered ISU strategies provided similar results and were better correlated to microscopy than OptiClust. It is important not to have a too high bootstrap: for biomonitoring purpose, it is better to determine neigbor species (which have similar ecologies) than not being able to determine it.





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#### Bias

- Physiological status of the cell / dead frustules

The amount of DNA depends on the status of the cells





Taxon	frustule %	status	Transfo / biovolume
Achnanthidium minutissimum	75	~	40
Amphora pediculus	13	X	0
Navicula cryptotenella	10	V	50
Melosira varians	2	V	10

#### Bias

**Completness of the reference database** 

For some species: no barcodes in the library



Taxon	frustule %	Barcode library	Transfo / biovolume
Achnanthidium minutissimum	75	No	0
Amphora pediculus	13	Yes	52
Navicula cryptotenella	10	Yes	40
Melosira varians	2	Yes	8

#### Bias

#### Think about neighbor/sister species



Taxon	frustule %
Encyonema minutum	75
Amphora pediculus	13
Navicula cryptotenella	10
Melosira varians	2

	N/A		
Taxon	Transfo / biovolum e		
Encyonema silesiacum	75		
Amphora nd	13		
Navicula cryptotenelloides	10		
Melosira varians	2		



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